

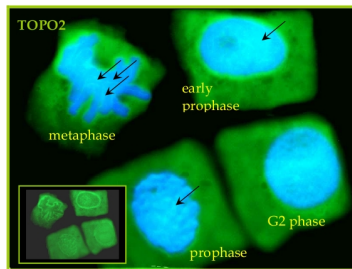
Product no **AS09 633****Goat anti-Rabbit IgG (H&L), DyLight® 488 conjugated****Product information**

Immunogen	Purified Rabbit IgG, whole molecule
Host	Goat
Clonality	Polyclonal
Purity	Immunogen affinity purified goat IgG.
Format	Lyophilized
Quantity	1 mg
Reconstitution	For reconstitution add 1.1 ml of sterile water. Let it stand 30 minutes at room temperature to dissolve. Prepare fresh working dilutions daily
Storage	Store lyophilized material at 2-8°C. Product is stable for 4 weeks at 2-8°C after rehydration. For long time storage after reconstitution, dilute the antibody solution with glycerol to a final concentration of 50% glycerol and store as liquid at -20°C, to prevent loss of enzymatic activity. For example, if you have reconstituted 1 mg of antibody in 1.1 ml of sterile water add 1.1 ml of glycerol. Such solution will not freeze in -20°C, If you are using a 1:5000 dilution prior to diluting with glycerol, then you would need to use a 1:2500 dilution after adding glycerol. Prepare working dilution prior to use and then discard. Be sure to mix well but without foaming.
Additional information	Concentration: 1.0 mg/ml Conjugate is present in 10 mM Sodium Phosphate, 0.15 M Sodium Chloride, pH 7.2, 1 % (w/v) BSA, Protease/IgG free. 0.05 % (w/v) sodium azide is added as preservative. DyLight® 488 has a maximum absorbance at 493 nm; Emax = 518 nm.

Application information

Recommended dilution	1 : 50- 1 : 5 000 (ICC), 1 : 20- 1 : 2000 (IHC), 1 : 3000 (IF)
Confirmed reactivity	Rabbit IgG heavy and light chains (H&L)
Predicted reactivity	Rabbit IgG Heavy and Light chains (H&L)
Additional information	Based in immunoelectrophoresis, this antibody reacts with heavy chains on rabbit IgG and light chains on all rabbit immunoglobulins. No reactivity is observed to non-immunoglobulin rabbit serum proteins based in immunoelectrophoresis. Purity of this antibody is > 95% based on SDS-PAGE.
Selected references	Burchardt et al. (2024) . Exploring the response of yellow lupine (<i>Lupinus luteus</i> L.) root to drought mediated by pathways related to phytohormones, lipid, and redox homeostasis. <i>BMC Plant Biol.</i> 2024 Nov 6;24(1):1049. doi: 10.1186/s12870-024-05748-4. Kamińska et al. (2024) . Comprehensive elucidation of the differential physiological kale response to cytokinins under in vitro conditions. <i>BMC Plant Biol.</i> 2024 Jul 15;24(1):674. doi: 10.1186/s12870-024-05396-8. Kucko et al. (2022) The acceleration of yellow lupine flower abscission by jasmonates is accompanied by lipid-related events in abscission zone cells, <i>Plant Science</i> , Volume 316, 2022,111173, ISSN 0168-9452, https://doi.org/10.1016/j.plantsci.2021.111173 . Namyslov et al. (2020) . Exodermis and Endodermis Respond to Nutrient Deficiency in Nutrient-Specific and Localized Manner. <i>Plants (Basel)</i> . 2020 Feb 6;9(2). pii: E201. doi: 10.3390/plants9020201. (immunolocalization) Fizesan et al. (2018) . Responsiveness assessment of a 3D tetra-culture alveolar model exposed to diesel exhaust particulate matter. <i>Toxicol In Vitro</i> . 2018 Aug 3;53:67-79. doi: 10.1016/j.tiv.2018.07.019. Liu et al. (2016) . Fold formation at the compartment boundary of <i>Drosophila</i> wing requires Yki signaling to suppress JNK dependent apoptosis. <i>Sci Rep</i> . 2016 Nov 29;6:38003. doi: 10.1038/srep38003. Wang et al. (2016) . Complementary expression of optomotor-blind and the Iroquois complex promotes fold formation to separate wing notum and hinge territories. <i>Dev Biol</i> . 2016 Aug 1;416(1):225-34. doi: 10.1016/j.ydbio.2016.05.020. Epub 2016 May 19

Application example



Seeds of field bean (*Vicia faba* L. subsp. minor var. Nadwiślański; DANKO Group; Sobiejuchy) were sterilized using sodium hypochlorite (0.3% v/v) and germinated in Petri dishes on wetted filter paper at room temperature. At 4 d after imbibition, dark-grown seedlings with primary roots 25 ± 5 mm long were selected for experiments. During incubations roots were oriented horizontally in a humid chamber and aerated continuously on a rotary water-bath shaker (30 rpm) at 23 °C. **Immunocytochemical assays** were performed according to the method prescribed earlier (Rybaczek and Maszewski 2006). Excised apical parts of roots (1.5 mm long) were fixed for 45 min (18 °C) in PBS-buffered 3.7% paraformaldehyde, washed several times with PBS and placed in a citric acid-buffered digestion solution (pH 5.0; 37 °C for 45 min) containing 2.5% pectinase (Fluka), 2.5% cellulase (Onozuka R-10; Serva) and 2.5% pectoliase (ICN). After removing the digestion solution, root tips were washed 3 times in PBS, rinsed with distilled water and squashed onto Super Frost Plus glass slides (Menzel-Gläser). Air-dried slides were pretreated with PBS-buffered 5% BSA at 20 °C for 50 min and incubated overnight in a humidified atmosphere (4 °C) with rabbit antibody raised against TOPO2 (Agrisera), dissolved in PBS containing 1% BSA (at a dilution of 1:500). Following incubation, slides were washed 3 times with PBS and incubated for 1 h (18 °C) with secondary goat anti-rabbit IgG DyLight®488 antibody (Agrisera, [AS09_633](#), 1:3000). Nuclear DNA was stained with 4',6-diamidino-2-phenyl-indole (DAPI, 0.4 µg/ml; Sigma-Aldrich). Following washing with PBS, slides were air dried and embedded in Vectashield Mounting Media for Fluorescence (Vector Laboratories). Observations were made using Optiphot-2 fluorescence microscope (Nikon) equipped with B-2A filter (blue light; 495 nm) for DyLight-conjugated antibodies and UV-2A filter (UV light; 365 nm) for DAPI. All images were recorded at exactly the same time of integration using DXM 1200 CCD camera.

Courtesy Dr. Dorota Rybaczek, Lodz University, Poland