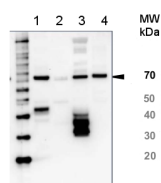


Product no **AS05 083****Anti-HSP70/HSC70 | Heat shock protein 70/Heat shock cognate protein 70 (serum)****Product information**

Immunogen	KLH-conjugated synthetic peptide conserved across all known sequences of HSP70 P08107 and HSC70 proteins P11142
Host	Rabbit
Clonality	Polyclonal
Purity	Total IgG. Protein G purified in PBS pH 7.4.
Format	Lyophilized
Quantity	100 µl
Reconstitution	For reconstitution add 100 µl of sterile water
Storage	Store lyophilized/reconstituted at -20°C; once reconstituted make aliquots to avoid repeated freeze-thaw cycles. Please remember to spin the tubes briefly prior to opening them to avoid any losses that might occur from material adhering to the cap or sides of the tube.
Additional information	For detection of plant and algal cytoplasmic hsp70 we recommend following product: AS08 371 .

Application information

Recommended dilution	1 : 1000 (IP), 1 : 1000-1: 5000 (WB)
Expected apparent MW	70 kDa
Confirmed reactivity	Fish, mammals, fungi: <i>Antrodia infirma</i> , <i>A. sinuosa</i> , <i>A. xantha</i> , <i>Catostomus commersonii</i> , <i>Gloeophyllum protractum</i> , <i>Gloeophyllum sepiarium</i> , <i>G. carbonarium</i> , <i>Junghunia luteoalba</i> , <i>Oligoporus sericiomollis</i> , <i>Phlebia cornea</i>
Predicted reactivity	Bovine, <i>Drosophila melanogaster</i> , Hen, Mouse, Rat
Not reactive in	No confirmed exceptions from predicted reactivity are currently known
Additional information	This antibody is recognizing both, the inducible and the constitutive Hsp70
Selected references	MacLellan et al. (2015). Chaperone roles for TMAO and HSP70 during hyposmotic stress in the spiny dogfish shark (<i>Squalus acanthias</i>). <i>J Comp Physiol B</i> . 2015 Jun 7. Bessemmer et al. (2014). Cardiorespiratory toxicity of environmentally relevant zinc oxide nanoparticles in the freshwater fish <i>Catostomus commersonii</i> . <i>Nanotoxicology</i> . 2014 Nov 27:1-10. Gorovits et al. (2013). Recruitment of the host plant heat shock protein 70 by tomato yellow leaf curl virus coat protein is required for virus infection. <i>PLoS One</i> , July 23;8(7).

Application example

10 µg of total protein from (1) killifish muscle, (2) bovine muscle, (3) chicken muscle, (4) rat liver, extracted with Protein Extraction Buffer, PEB ([AS08 300](#)) and separated on 4-12% NuPage (Invitrogen) LDS-PAGE and blotted 1h to PVDF. Blots were blocked in 5 % non-fat milk for 1h at room temperature with agitation. Blots were incubated in the primary antibody at a dilution of 1 : 5000 (in blocking reagent) for 1h at room temperature with agitation. The antibody solution was decanted and the blot was rinsed briefly twice, then washed once for 15 min and 3 times for 5 min in TBS-T at room temperature with agitation. Blots were incubated in secondary antibody (Agrisera anti-rabbit IgG horse radish peroxidase conjugated, [AS09 602](#)) diluted to 1:25 000 in blocking reagent for 1h at room temperature with agitation. The blots were washed as above and developed for 5 min with chemiluminescent detection reagent, according to the manufacturers instructions. Images of the blots were obtained using a CCD imager (FluorSMax, Bio-Rad) and Quantity One software (Bio-Rad). Exposure time was 5 min.