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## Product no AS13 2705

## Lhcb2-P | LHCII type II chlorophyll a/b-binding protein, phosphorylated

## **Product information**

Immunogen KLH-conjugated synthetic peptide: RRT\*VKSTPQS, where T\* indicates phospho-Thr

**Host** Rabbit

Clonality Polyclonal

**Purity** Immunogen affinity purified serum in PBS pH 7.4.

Format Lyophilized

Quantity 25 µg

**Reconstitution** For reconstitution add 25 μl of sterile water

Store lyophilized/reconstituted at -20°C; once reconstituted make aliquots to avoid repeated freeze-thaw cycles. Please

remember to spin the tubes briefly prior to opening them to avoid any losses that might occur from material adhering to the cap or sides of the tube.

Application information

Recommended dilution 1:10 000 (WB)

Expected | apparent

25 | 25 kDa for Arabidopsis thaliana

Predicted reactivity

Arachis hypogaea, Colobanthus quitensis Kunt Bartl, Hordeum vulgare, Mesembryanthemum crystallinum, Nicotiana tabacum, Pisum sativum, Phaseolus vulgaris, Solanum lycopersicum, Spinacia oleracea, Physcomitrium patens Species of your interest not listed? Contact us

Not reactive in

No confirmed exceptions from predicted reactivity are currently known

Selected references

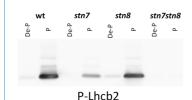
Wu et al. (2021). Formation of light-harvesting complex (LHC) II aggregates from LHCII-PSI-LHCI complexes in rice plants under high light. J Exp Bot. 2021 May 3:erab188. doi: 10.1093/jxb/erab188. Epub ahead of print. PMID: 33939808

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Bychkov et al. (2019). Melatonin modifies the expression of the genes for nuclear- and plastid-encoded chloroplast proteins in detached Arabidopsis leaves exposed to photooxidative stress. Plant Physiology and Biochemistry, doi.org/10.1016/j.plaphy.2019.10.013.

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## Application example



1 μg of thylakoid membranes isolated from *Arabidopsis thaliana* wild-type and respective mutants were solubilized with 3X LB (6 M urea, 12% SDS, 30% glycerol, 100 mM DTT, 150 mM Tris pH7.0, 0.8% Comassie G-250). 1 µg of total chlorophyll was loaded and separated on 16% SDS-PAGE, and then blotted for 2 h onto nitrocellulose membrane. Blots were blocked with milk powder for 2 h and then incubated in the primary antibody solution (AS13 2705, 1: 5 000) for 2.5 h, which was then decanted and the blot was washed 3 times for 5 min in TBST. Membrane was incubated in secondary



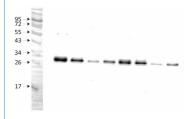
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antibody (anti-rabbit IgG horse radish peroxidase conjugated) diluted to 1:10 000 in for 1 h, followed by washing steps as above. All the steps fallowing transfer were performed in room temperature (RT) with agitation. Membrane was developed for 5 min with ECL according to the manufacturer's instructions and recorded using FujiFilm CCD camera with 30 s increment time for around 5 min.

Courtesy of a phd candidate Małgorzata Pietrzykowska, Umeå Plant Science Centre, Sweden



1.0 µg of chlorophyll from mesophyll (M) and bundle sheath (BS) thylakoids of various treatments of *Zea mays* extracted with 0.4 M sorbitol, 50 mM Hepes NaOH, pH 7.8, 10 mM NaCl, 5 mM MgCl<sub>2</sub> and 2 mM EDTA. Samples were denatured with Laemmli buffer at 75 °C for 5 min and were separated on 12% SDS-PAGE and blotted 30 min to PVDF using wet transfer. Blot was blocked with 5% BSA for 1h at room temperature (RT) with agitation. Blot was incubated in the primary antibody at a dilution of 1: 10 000 overnight at 4 °C with agitation in 1% BSA in TBS-T. The antibody solution was decanted and the blot was washed 4 times for 5 min in TBS-T at RT with agitation. Blot was incubated in secondary antibody (anti-rabbit IgG horse radish peroxidase conjugated, from Agrisera, <u>AS09 602</u>) diluted to 1:25 000 in 1 % BSA in TBS-T for 1h at RT with agitation. The blot was washed 5 times for 5 min in TBS-T and 2 times for 5 min in TBS, and developed for 1 min with 1.25 mM luminol, 0.198 mM coumaric acid and 0.009% H<sub>2</sub>O<sub>2</sub> in 0.1 M Tris- HCl, pH 8.5. Exposure time in ChemiDoc System was 54 seconds.

Courtesy Dr. Wiola Wasilewska, Faculty of Biology, University of Warsaw, Poland