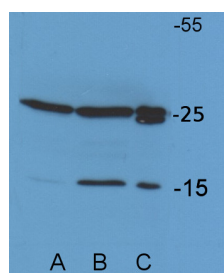


Product no **AS09 407****Anti-Lhcb5 | CP26 (Lhcb5) homolog, Chlamydomonas****Product information**

| | |
|-----------------------|---|
| Immunogen | KLH-conjugated synthetic peptide derived from <i>Chlamydomonas reinhardtii</i> Lhcb5 protein sequence, UniProt: Q9FEK6 |
| Host | Rabbit |
| Clonality | Polyclonal |
| Purity | Serum |
| Format | Lyophilized |
| Quantity | 50 µl |
| Reconstitution | For reconstitution add 50 µl of sterile water |
| Storage | Store lyophilized/reconstituted at -20°C; once reconstituted make aliquots to avoid repeated freeze-thaw cycles. Please remember to spin the tubes briefly prior to opening them to avoid any losses that might occur from material adhering to the cap or sides of the tube. |

Application information

| | |
|-------------------------------|---|
| Recommended dilution | 1 : 5000-1 : 10 000 (WB) |
| Expected apparent MW | 30 26 kDa |
| Confirmed reactivity | <i>Chlamydomonas reinhardtii</i> |
| Predicted reactivity | <i>Chlamydomonas reinhardtii</i> |
| Not reactive in | Other algae |
| Additional information | For optimal detection, load/well should be from 5 to 10 µg of total protein. Higher protein load combined with weak blocking will contribute to detection of other bands. |
| Selected references | <p>Kim et al. (2024). Photoautotrophic cultivation of a <i>Chlamydomonas reinhardtii</i> mutant with zeaxanthin as the sole xanthophyll. <i>Biotechnol Biofuels Bioprod.</i> 2024 Mar 14;17(1):41. doi: 10.1186/s13068-024-02483-8.</p> <p>Cecchin et al (2021) LPA2 protein is involved in photosystem II assembly in <i>Chlamydomonas reinhardtii</i>. <i>Plant J.</i> 2021 Jul 4. doi: 10.1111/tpj.15405. Epub ahead of print. PMID: 34218480.</p> <p>Gonzaga Heredia-Martinez et al. (2018). Chloroplast damage induced by the inhibition of fatty acid synthesis triggers autophagy in <i>Chlamydomonas</i>. <i>Plant Physiol.</i> Sept. 2018.</p> <p>Correa-Galvis et al. (2016). Photosystem II Subunit PsbS Is Involved in the Induction of LHCSR Protein-dependent Energy Dissipation in <i>Chlamydomonas reinhardtii</i>. <i>J Biol Chem.</i> 2016 Aug 12;291(33):17478-87. doi: 10.1074/jbc.M116.737312.</p> <p>Muranaka et al. (2015). TEF30 interacts with photosystem II monomers and is involved in the repair of photodamaged photosystem II in <i>Chlamydomonas reinhardtii</i>. <i>Plant Physiol.</i> 2015 Dec 7. pii: pp.01458.2015.</p> <p>Drop et. al (2014). Consequences of state transitions on the structural and functional organization of Photosystem I in the green alga <i>Chlamydomonas reinhardtii</i>. <i>Plant J.</i> 2014 Feb 8. doi: 10.1111/tpj.12459.</p> |

Application example

Chlamydomonas reinhardtii membrane extract (**A**), *Chlamydomonas reinhardtii* total cell extract, prepared by sonication, loading 14 µl equivalent to 30 µg of total protein (**B**), *Chlamydomonas reinhardtii* total cell extract, rapid, prepared directly by spinning down the cells and lysis of cell pellet in SDS-PAGE sample buffer and loading 14 µl equivalent to 98 µg of total protein (**C**), denatured at 100°C for 5 min. were separated on 15 % SDS-PAGE and blotted 1h to PVDF using tank transfer. Blots were blocked with 1 % blocking buffer (2 ml blocking reagent stock solution ROCHE 11 520 709 001 in 20 ml TBS) for ON at 4°C without agitation. Blot was incubated in the primary antibody at a dilution of 1: 25 000 for 1h at RT with agitation. The antibody solution was decanted and the blot was rinsed briefly twice, then washed once for 15 min and 3 times for 5 min in TBS-T at RT with agitation. Blot was incubated in secondary antibody (anti-rabbit IgG horse radish peroxidase conjugated, from Agrisera [AS09](#)

This product is **for research use only** (not for diagnostic or therapeutic use)

contact: support@agrisera.com

Agrisera AB | Box 57 | SE-91121 Vännäs | Sweden | +46 (0)935 33 000 | www.agrisera.com

602) diluted to 1:20 000 in for 1h at RT with agitation. The blot was washed as above and developed according to manufacture instructions. Exposure time was 8 seconds.

Courtesy of Nadine Coosemans, Laboratoire de génétique et physiologie des microalgues, Université de Liège, Belgium