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### Product no AS06 143

# Anti-PsaH | PSI-H subunit of photosystem I, Chlamydomonas

#### **Product information**

Immunogen Recombinant PsaH protein from Chlamydomonas reinhardtii P13352

**Host** Rabbit

Clonality Polyclonal

Purity Serum

Format Lyophilized

Quantity 200 μl

**Reconstitution** For reconstitution add 200 μl of sterile water

Store lyophilized/reconstituted at -20°C; once reconstituted make aliquots to avoid repeated freeze-thaw cycles. Please remember to spin the tubes briefly prior to opening them to avoid any losses that might occur from material adhering to the cap or sides of the tube.

## Application information

Recommended dilution 1:10 000 (WB)

Expected | apparent

10 | 10 for Chlamydomonas reinhardtii

Confirmed reactivity Arabidopsis thaliana (weak), Chlamydomonas reinhardtii, Hordeum vulgare

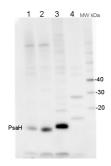
**Predicted reactivity** Species of your interest not listed? Contact us

Not reactive in Synechococcus sp. PCC 7942

Selected references

Nama et al. (2018). Non-photochemical quenching-dependent acclimation and thylakoid organization of Chlamydomonas reinhardtii to high light stress. Photosynth Res. 2018 Jul 7. doi: 10.1007/s11120-018-0551-7. Winck (2011). Nuclear proteomics and transcription factor profiling. Dissertation, University of Posdam.

## Application example



2 µg of total leaf protein of Arabidopsis thaliana (1) and Hordeum vulgare (2) and total cellular protein of Chlamydomonas reinhardtii (3) and Synechococcus PCC 7942 (4) isolated with Agrisera Protein Extraction Buffer (AS08 300), were separated on 4-12% Nupage Bis-Tris gels in in MES running buffer (Invitrogen) at 200V for 35 minutes. Proteins were transferred for 80 minutes at 30V to a PVDF membrane pre-wetted in methanol and equilibrated in 1X transfer buffer. Blots were blocked immediately following transfer in 2% blocking reagent in 20 mM Tris, 137 mM sodium chloride pH 7.6 with 0.1% (v/v) Tween-20 (TBS-T) and probed with anti-PsaH (AS06 143, 1:10000) and secondary HRP-conjugated goat anti-rabbit antibody (1:50 000) for 1 hr in TBS-T containing 2% blocking reagent. Antibody incubations were followed by washings in TBS-T (15, +5, +5, +5 min). All steps were performed at RT with agitation. Signals was detected after 3 s using chemiluminescent detection reagent (GE Healthcare) according to the manufacturers instructions and a CCD imager (FluorSMax, Bio-Rad).