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**Product no** AS15 3098

# Anti-RDR6 | RNA-dependent RNA polymerase 6

## Product information

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|---|---|
| **Immunogen** | KLH-conjugated peptide derived from *Arabidopsis thaliana* RDR6 sequence, Uniprot: Q9SG02 ,TAIR: At3g49500 |
| **Host** | Rabbit |
| **Clonality** | Polyclonal |
| **Purity** | Immunogen affinity purified serum in PBS pH 7.4. |
| **Format** | Lyophilized |
| **Quantity** | 50 µg |
| **Reconstitution** | For reconstitution add 50 µl of sterile water |
| **Storage** | Store lyophilized/reconstituted at -20°C; once reconstituted make aliquots to avoid repeated freeze-thaw cycles. Please remember to spin the tubes briefly prior to opening them to avoid any losses that might occur from material adhering to the cap or sides of the tube. |

## Application information

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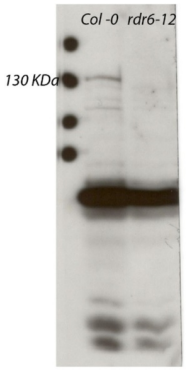
| **Recommended dilution** | 1 : 2000-1 : 6000 (WB) |
| **Expected \| apparent MW** | 136.9 \| 130 kDa |
| **Confirmed reactivity** | *Arabidopsis thaliana* |
| **Not reactive in** | *Nicotiana tabacum, Solanum lycopersicum, Zea mays* |

**Application example**




50 µg of total protein from *Arabidopsis thaliana* whole vegetative rosette wild type Col-0 **(a)** *rdr6-12* mutant **(b)** extracted with extraction buffer (50 mM Tris pH 7.5; 150 mM NaCl; 1 mM EDTA; 10 % v/v Glycerin; 1 mM DTT, 1x Complete Protease Inhibitor Cocktail, Roche) and denatured with Laemmli buffer at 95°C/5 min., were separated on 7.5 % SDS-PAGE and blotted 1.5 h to PVDF using tank transfer. Blots were blocked with blocking buffer (5% milk powder; 1x TBS; 0.1% Tween-20) overnight at 4°C with agitation. Blot was incubated in the primary antibody at a dilution of 1:6000 for 2h at RT with agitation. The antibody solution was decanted and the blot was rinsed briefly and then washed tree times for 15 min in TBS-T at RT with agitation. Blot was incubated in secondary antibody (anti-rabbit IgG horse radish peroxidase conjugated, from Agrisera, AS09 602) diluted to 1:20 000 in blocking buffer for 1h at RT with agitation. The blot was washed as above and developed for 5 min with chemiluminescent detection reagent and expose to Amersham Hyperfilms ECL for 3 minutes.

Courtesy of Dr. Pablo Manavella, Instituto de Agrobiotecnología del Litoral (IAL), Argentina