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Product no AS15 2830

## Anti-SUS1 | Sucrose synthase 1

## **Product information**

Immunogen His-tagged, full length Arabidopsis thaliana SUS1, UniProt: P49040, TAIR: AT5G20830

Host Rabbit

Clonality Polyclonal

Purity Serum

Format Lyophilized

Quantity 50 μl

**Reconstitution** For reconstitution add 50 μl of sterile water

Store lyophilized/reconstituted at -20°C; once reconstituted make aliquots to avoid repeated freeze-thaw cycles. Please remember to spin the tubes briefly prior to opening them to avoid any losses that might occur from material adhering to

the cap or sides of the tube.

## Application information

Recommended dilution 1:10 000 (WB)

Expected | apparent

Confirmed reactivity Arabidopsis thaliana, Hordeum vulgare, Miscanthus x giganteus, Olea europea, Pinus yunnanensis, Zea mays

93 kDa

Predicted reactivity

Brassica sp., Glycine max, Gossypium sp., Hevea brasiliensis, Jatropha curas, Mangifera indica, Manihot esculenta,

Theobroma cacao, Pisum sativum, populus tomentosa, Ricinus communis

Species of your interest not listed? Contact us

Not reactive in No confirmed exceptions from predicted reactivity are currently known

Selected references

Bilska-Kos et al. (2020). Sucrose phosphate synthase (SPS), sucrose synthase (SUS) and their products in the leaves of Miscanthus× giganteus and Zea mays at low temperature. Planta . 2020 Jul 16;252(2):23. doi:

10.1007/s00425-020-03421-2.

Kleczkowski LA & Decker DD (2015) Sugar activation for production of nucleotide sugars as substrates for glycosyltransferases in plants. J. Appl. Glycosci. (in press).

## **Application example**



10 μg of total protein from Arabidopsis thaliana leaf (1), Hordeum vulgare leaf (2), Zea mays leaf (3) were extracted with Protein Extraction Buffer PEB (AS08 300). Samples were diluted with 1X sample buffer (NuPAGE LDS sample buffer (Invitrogen) supplemented with 50 mM DTT and heat at 70°C for 5 min and keept on ice before loading. Protein samples were separated on 4-12% Bolt Plus gels, LDS-PAGE and blotted for 70 minutes to PVDF using tank transfer. Blots were blocked immediately following transfer in 2% blocking reagent or 5% non-fat milk dissolved in 20 mM Tris, 137 mM sodium chloride pH 7.6 with 0.1% (v/v) Tween-20 (TBS-T) for 1h at room temperature with agitation. Blots were incubated in the primary antibody at a dilution of 1: 10 000 (in blocking reagent) for 1h at room temperature with agitation. The antibody solution was decanted and the blot was rinsed briefly twice, and then washed 1x15 min and 3x5 min with TBS-T at room temperature with agitation. Blots were incubated in secondary antibody (anti-rabbit IgG horse radish peroxidase conjugated, recommended secondary antibody AS09 602, Agrisera) diluted to 1:25 000 in blocking reagent for 1h at room temperature with agitation. The blots were washed as above. The blot was developed for 5 min with detection reagent according the manufacturers instructions. Images of the blots were obtained using a CCD imager (VersaDoc MP 4000) and Quantity One software (Bio-Rad). Exposure time was 1 minute.