

This product is for research use only (not for diagnostic or therapeutic use)

contact: support@agrisera.com

Agrisera AB | Box 57 | SE-91121 Vännäs | Sweden | +46 (0)935 33 000 | www.agrisera.com

Product no AS19 4311

PGRL1 | PGR5-like protein 1A (chloroplastic)

Product information

Immunogen KLH-conjugated peptide derived from Arabidopsis thaliana PGRL1A UniProt: Q8H112, TAIR: At4g22890

Peptide used to elicit this antibody is conserved in both isoforms PGRL1A and 1B of Zea mays

Host Rabbit

Clonality Polyclonal

Purity Immunogen affinity purified serum in PBS pH 7.4.

Format Lyophilized

Quantity 50 μg

Reconstitution For reconstitution add 50 μl of sterile water

Storage Storage Store lyophilized/reconstituted at -20 °C; once reconstituted make aliquots to avoid repeated freeze-thaw cycles. Please remember to spin the tubes briefly prior to opening them to avoid any losses that might occur from material adhering to

the cap or sides of the tube.

Application information

Recommended dilution 1:1000 (WB)

Expected | apparent

29 | 29 kDa (Arabidopsis thaliana, Spinacia oleracea)

Predicted reactivity Dichanthelium oligosanthes, Glycine soja, Gossypium arboreum, Klebsormidium nitens, Nelumbo nucifera, Noccaea

caerulescens, Physcomitrium patens, Prunus dulcis, Prunus yedoensis, Rhizophora mucronatamm, Solanum

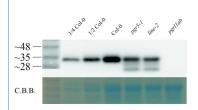
chacoense, Trifolium medium, Zea mays, Vitis vinifera Species of your interest not listed? <u>Contact us</u>

Not reactive in Chlorella sp., diatoms

Additional information This product can be sold containing ProClin if requested

Selected references McKinnon et al. (2020). Membrane Chaperoning of a Thylakoid Protease Whose Structural Stability Is Modified by the

Protonmotive Force. Plant Cell DOI: 10.1105/tpc.19.00797



Whole leaf protein (corresponding to 3 mg fresh weight) from the *Arabidopsis thaliana* lines Columbia-0 (Col-0), PGR5 knock out (pgr5-1), line-2 (line-2) and PGRL1A+B knock out (pgr11ab) was extracted and denatured with 2x Tricine sample buffer (100mM Tris/HCl, 24% (w/v) glycerol, 8% SDS and 15mM DTT) by heating for 5 min at 70°C. The samples were separated on a 10% Tris Tricine gel and blotted overnight on PVDF-membrane using capillary blotting. The membrane was blocked with 5% milk in Tris-Buffered saline and 0.1% Tween (TBS-T (50mM Tris, 150mM NaCl, 0.1% Tween, pH 7.5)) for 1h at room temperature with agitation and subsequently incubated in the primary antibody at a dilution of 1:5000 overnight at 4°C with agitation in 1% milk in TBS-T. The antibody solution was decanted and the blot was rinsed briefly, then washed 3 times for 10 min in TBS-T at RT with agitation. The membrane was incubated in secondary antibody (anti-rabbit IgG horse radish peroxidase conjugated, from Agrisera, AS09 602) diluted to 1:10 000 in 1% milk in TBS-T for 1h at RT with agitation. The blot was washed 3 times for 10 min in TBS-T, once for 5 min in TBS (50mM Tris, 150mM NaCl, pH 7.5) and developed for 5 min with chemiluminescent detection reagent according to manufacture's recommendations. Exposure time was 5 seconds.

Courtesy of Jan-Ferdinand Penzler from Ludwig-Maximilians-University Munich (LMU), Dario Leister Lab, Germany

Experimental conditions for detection in extracts of Picea abies and Pinus sylvestris:



This product is for research use only (not for diagnostic or therapeutic use)

contact: support@agrisera.com

Agrisera AB | Box 57 | SE-91121 Vännäs | Sweden | +46 (0)935 33 000 | www.agrisera.com

25 μg/well of total protein extracted from needles of *Picea abies* and *Pinus sylvestris*. Exact buffer components were: 10% trichloroacetic acid (TCA)/ acetone buffer. Then followed by a methanol buffer (80% methanol with 0.1 M ammonium acetate) wash and an 80% acetone wash step. Subsequently, total of 1.8 ml mixture of 1:1 phenol (pH 8.0)/SDS buffer (30% sucrose, 2% SDS, 0.1M Tris-HCl, pH 8.0, 5% 2-mercaptoethanol) were added to the tube to extract proteins from dry pellets. After centrifuging at 16,000 g, 4 °C for 3 min, 0.4 ml phenol phase extraction were mixed with 1.6 ml methanol buffer and then centrifuged at 16,000 g, 4 °C for 3 min. The pellets were washed twice with methanol and 80% acetone. Final protein pellets were dissolved in 0.2 ml Laemmli sample buffer, and denatured at 100 °C for 10 min. Samples were separated in the cold on 12 % SDS-PAGE and blotted for 1 h to PVDF membrane, using: wet transfer in the cold. Blot was blocked with 5 % milk for: 1h/RT with agitation. Blot was incubated in the primary antibody at a dilution of 1: 1000 for ON/4 °C with agitation. The antibody solution was decanted and the blot was rinsed briefly twice, then washed once for 15 min and 3 times for 5 min in TBS-T at RT with agitation. Blot was incubated in matching secondary antibody (anti-rabbit IgG horse radish peroxidase conjugated) diluted to 1: 10 000 in for 1h/RT with agitation. The blot was washed as above and developed with a following chemiluminescent detection reagent:. Exposure time was 1-5 minutes. <u>Yang</u> et al. 2020.