

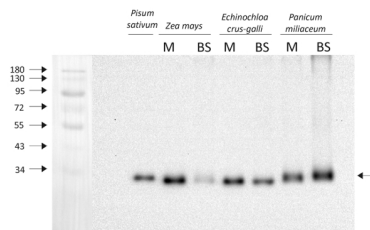
Product no **AS06 142-33****PsbO | 33 kDa of the oxygen evolving complex (OEC) of PSII (anti-protein)****Product information**

<b>Immunogen</b>	Native purified 33 kDa protein from <i>Spinacia oleracea</i> UniProt: <a href="#">P12359</a>
<b>Host</b>	Rabbit
<b>Clonality</b>	Polyclonal
<b>Purity</b>	Total IgG. Protein G purified in PBS pH 7.4.
<b>Format</b>	Lyophilized
<b>Quantity</b>	50 µl
<b>Reconstitution</b>	For reconstitution add 50 µl of sterile water
<b>Storage</b>	Store lyophilized/reconstituted at -20°C; once reconstituted make aliquots to avoid repeated freeze-thaw cycles. Please remember to spin the tubes briefly prior to opening them to avoid any losses that might occur from material adhering to the cap or sides of the tube.
<b>Additional information</b>	Total IgG fraction has been purified by 40% ammonium sulphate precipitation followed by DEAE cellulose chromatography  This product can be sold containing ProClin if requested

**Application information**

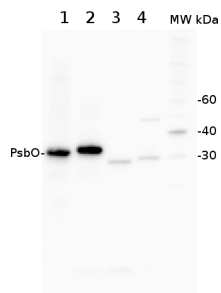
<b>Recommended dilution</b>	1 : 2000-1 : 5000 (WB)
<b>Expected   apparent MW</b>	35   33 kDa
<b>Confirmed reactivity</b>	<i>Arabidopsis thaliana</i> , <i>Chlamydomonas reinhardtii</i> , <i>Chlorella</i> sp. DT, <i>Chlorella vulgaris</i> , <i>Echinochloa crus-galli</i> , <i>Eucalyptus grandis</i> x <i>Eucalyptus camaldulensis</i> , <i>Festuca arundinacea</i> cv. Kord, <i>Haematococcus pluvialis</i> , <i>Hordeum spontaneum</i> , <i>Hordeum vulgare</i> , <i>Nicotiana benthamiana</i> , <i>Oryza sativa</i> , <i>Panicum miliaceum</i> , <i>Picea abies</i> , <i>Pinus banksiana</i> , <i>Pisum sativum</i> , <i>Spinacia oleracea</i> , <i>Solanum tuberosum</i> cultivar Taedong Valley, <i>Synechococcus</i> sp. PCC7002, sp. PCC 7942 and sp. PCC6803, <i>Thellungiella salsuginea</i> , <i>Triticum aestivum</i> , <i>Zea mays</i>
<b>Predicted reactivity</b>	<i>Glycine max</i> , <i>Nicotiana tabacum</i> , <i>Solanum lycopersicum</i> , <i>Populus trichocarpa</i> , <i>Zostera marina</i>  Species of your interest not listed? <a href="#">Contact us</a>
<b>Not reactive in</b>	No confirmed exceptions from predicted reactivity are currently known
<b>Additional information</b>	This antibody can be used as a loading control for <i>Chlamydomonas reinhardtii</i> while it not so suitable for higher plants as accumulation of these proteins might drop to 12.5-25 % of the WT level in mutants defective for PSII core ( <a href="#">Schult</a> et al. 2007).
<b>Selected references</b>	<a href="#">Nagy</a> et al. (2023). Photoautotrophic and sustained H2 production by the pgr5 mutant of <i>Chlamydomonas reinhardtii</i> in simulated daily light conditions. International Journal of Hydrogen Energy Volume 53, 31 January 2024, Pages 760-769. <a href="#">Cecchin</a> et al (2021) LPA2 protein is involved in photosystem II assembly in <i>Chlamydomonas reinhardtii</i> . Plant J. 2021 Jul 4. doi: 10.1111/tpj.15405. Epub ahead of print. PMID: 34218480. <a href="#">Loudya</a> et al. (2021) Cellular and transcriptomic analyses reveal two-staged chloroplast biogenesis underpinning photosynthesis build-up in the wheat leaf. Genome Biol. 2021 May 11;22(1):151. doi: 10.1186/s13059-021-02366-3. PMID: 33975629; PMCID: PMC8111775. <a href="#">Terentyev</a> (2020: The Main Structural and Functional Characteristics of Photosystem-II-Enriched Membranes Isolated From Wild Type and <i>cia3</i> Mutant <i>Chlamydomonas reinhardtii</i> . Life (Basel). 2020 May 14;10(5):E63. doi: 10.3390/life10050063.. <a href="#">Tang</a> et al. (2020). OsNSUN2-Mediated 5-Methylcytosine mRNA Modification Enhances Rice Adaptation to High Temperature. Dev Cell. 2020 May 4;53(3):272-286.e7. doi: 10.1016/j.devcel.2020.03.009. <a href="#">Smythers</a> et al. (2019). Characterizing the effect of Poast on <i>Chlorella vulgaris</i> , a non-target organism. Chemosphere Volume 219, March 2019, Pages 704-712.

## Application example



1 µg of chlorophyll from *Pisum sativum* thylakoids and from mesophyll (M) and bundle sheath (BS) thylakoids of *Zea mays*, *Echinochloa crus-galli*, *Panicum miliaceum* extracted with 0.4 M sorbitol, 50 mM Hepes NaOH, pH 7.8, 10 mM NaCl, 5 mM MgCl<sub>2</sub> and 2 mM EDTA were loaded to lanes. Samples were denatured with Laemmli buffer at 75 °C for 5 min and were separated on 12% SDS-PAGE and blotted 30 min to PVDF using wet transfer. Blot was blocked with 5% milk for 2 h at room temperature (RT) with agitation. Blot was incubated in the primary antibody AS06 142-33 (Lot 1902) at a dilution of 1: 2000 overnight at 40°C with agitation in 1% milk in TBS-T. The antibody solution was decanted and the blot was washed 4 times for 5 min in TBS-T at RT with agitation. Blot was incubated in secondary antibody (anti-rabbit IgG HRP conjugated, from Agrisera, AS09 602, Lot 1902, as recommended) diluted to 1:20 000 in 1% milk in TBS-T for 1 h at RT with agitation. The blot was washed 5 times for 5 min in TBS-T and 2 times for 5 min in TBS, and developed for 1 min with 1.25 mM luminol, 0.198 mM coumaric acid and 0.009% H<sub>2</sub>O<sub>2</sub> in 0.1 M Tris-HCl, pH 8.5. Exposure time in ChemiDoc System was 100 seconds

Courtesy of Dr. Wioleta Wasilewska-Dębowska, Warsaw University, Poland



2 µg of total protein from *Arabidopsis thaliana* leaf (1), *Hordeum vulgare* leaf (2), *Chlamydomonas reinhardtii* total cell (3), *Synechococcus* sp. 7942 total cell (4), all extracted with PEB (AS08 300) were separated on 4-12% NuPage (Invitrogen) LDS-PAGE and blotted 1 h to PVDF. Blots were blocked immediately following transfer in 2% ECL Advance blocking reagent in 20 mM Tris, 137 mM sodium chloride pH 7.6 with 0.1% (v/v) Tween-20 (TBS-T) for 1 h at room temperature with agitation. Blots were incubated in the primary antibody at a dilution of 1: 10 000 for 1 h at room temperature with agitation. The antibody solution was decanted and the blot was rinsed briefly twice, then washed once for 15 min and 3 times for 5 min in TBS-T at room temperature with agitation. Blots were incubated in secondary antibody (anti-rabbit IgG horse radish peroxidase conjugated) diluted to 1:50 000 in 2% blocking solution for 1 h at room temperature with agitation. The blots were washed as above and developed for 5 min with chemiluminescent detection reagent according the manufacturers instructions. Images of the blots were obtained using a CCD imager (FluorSMax, Bio-Rad) and Quantity One software (Bio-Rad).

**Note:** Detection level in algal species can be improved by adjustment of western blot protocol. Please [inquire](#).