

Agrisera

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Product no **AS14 2775**

V-ATPase, B | vacuolar H⁺-ATPase subunit B

Product information

Background	V-ATPase , subunit B is a non-catalytic subunit of the peripheral V1 complex of vacuolar ATPase. Alternative names: Vacuolar proton pump subunit B1, vacuolar H ⁺ -ATPase subunit B, V-ATPase 57 kDa subunit.
Immunogen	<u>KLH</u> -conjugated synthetic peptide chosen from <i>Arabidopsis thaliana</i> V-ATPase subunit B, isoform B1: UniProt: Q683E8 , TAIR: AT1G76030 , isoform B2 UniProt: Q9SZN1 , TAIR: AT4G38510 , isoform B3: UniProt: Q8W4E2 , TAIR: AT1G20260
Host	Rabbit
Clonality	Polyclonal
Purity	Affinity purified serum in PBS, pH 7.4
Format	Lyophilized
Quantity	50 µg
Reconstitution	For reconstitution add 50 µl of sterile water.
Storage	Store lyophilized/reconstituted at -20°C; make aliquots to avoid repeated freeze-thaw cycles. Please, remember to spin tubes briefly prior to opening them to avoid any losses that might occur from material adhering to the cap or sides of the tubes.
Tested applications	Western blot (WB)
Related products	collection of antibodies to vacuolar proteins AS09 467 Anti-V-ATPase subunit A vacuolar H ⁺ -ATPase, rabbit antibodies AS09 468 Anti-V-ATPase subunit c vacuolar H ⁺ -ATPase, subunit c (16 kDa), rabbit antibodies AS09 497 Anti- V-ATPase subunit D V-type proton ATPase subunit D, rabbit antibodies AS07 213 Anti-V-ATPase subunit E of tonoplast H ⁺ ATPase, rabbit antibodies AS09 499 Anti-V-ATPase subunit H V-type proton ATPase subunit H, rabbit antibodies Plant protein extraction buffer
Additional information	Antibodies will detect target protein in a few µg of a crude preparation loaded per well. If purified preparations of vacuolar and plasma membranes are used, one µg load per well should be sufficient.

Application information

Recommended dilution	1 : 1000 (WB)
Expected apparent MW	53 57 kDa (<i>Vigna radiata</i>)
Confirmed reactivity	<i>Arabidopsis halleri</i> , <i>Nicotiana tabaccum</i> , <i>Theilungiella salsuginea</i> , <i>Vigna radiata</i>
Predicted reactivity	<i>Acetabularia sp.</i> , <i>Arundo donax</i> , <i>Chlamydomonas reinhardtii</i> , <i>Cucumis sativus</i> , <i>Glycine soja</i> , <i>Gossypium mexicanum</i> , <i>Halostachys caspica</i> , <i>Haloxylon ammodendron</i> , <i>Hordeum vulgare</i> , <i>Medicago truncatula</i> , <i>Mesembryanthemum crystallinum</i> , <i>Ostreococcus tauri</i> , <i>Oryza sativa</i> , <i>Panax ginseng</i> , <i>Physcomitrella patens</i> , <i>Pinus sylvestris</i> , <i>Populus trichocarpa</i> , <i>Pyrus sp.</i> , <i>Ricinus communis</i> , <i>Theobroma cacao</i> , <i>Triticum aestivum</i> , <i>Zea mays</i> , <i>Zostera marina</i> Species of your interest not listed? Contact us

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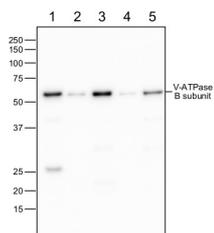
Not reactive in | No confirmed exceptions from predicted reactivity are currently known.

Additional information | Protein or membrane sample should be treated at 70°C for 10 min before loading on the gel.

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Application example



Proteins were separated by SDS-PAGE and transferred to an Immobilon-P membrane (Millipore) using Trans-Blot SD Semi-Dry Transfer Cell (Bio-Rad) with transfer buffer (100 mM Tris, 192 mM Glycine, 0.02% (w/v) SDS and 5% (v/v) methanol). After treatment with 1% blocking agent, the membrane filter was incubated with the primary antibody (1:1000) and then with horseradish peroxidase (HRP)-conjugated goat anti-rabbit IgG (H+L) (Agrisera [AS09 602](#), 1:25 000). Chemiluminescent reagent was used for detection of antigens. Chemiluminescence was detected with a Light-Capture II imaging device with a cooled CCD camera (Atto).

Samples:

- 1: 10 µg of 100,000 x g precipitate prepared from *Arabidopsis thaliana* 6 weeks old shoot.
- 2: 0.2 µg of vacuolar membrane enriched fraction prepared from *Arabidopsis thaliana* 6 weeks old shoot.
- 3: 2 µg of vacuolar membrane enriched fraction prepared from *Arabidopsis thaliana* 6 weeks old shoot.
- 4: 0.2 µg of vacuolar membrane enriched fraction prepared from *Vigna radiata* 4 days old hypocotyls. 5: 2 µg of vacuolar membrane enriched fraction prepared from *Vigna radiata* 4 days old hypocotyls.

Courtesy of Drs. Masayoshi Maeshima and Dr Shoji Segami, Nagoya University, Japan