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Agrisera AB | Box 57 | SE-91121 Vännäs | Sweden | +46 (0)935 33 000 | www.agrisera.com

Product no AS16 4030

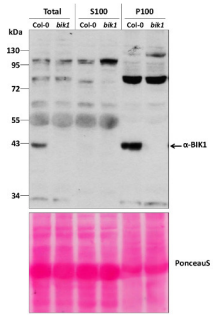
Anti-BIK1 | Botrytis-induced kinase 1

Product information

Immunogen	KLH-conjugated synthetic peptide derived from *Arabidopsis thaliana* BIK1 sequence, UniProt: O48814, TAIR: AT2G39660
Host	Rabbit
Clonality	Polyclonal
Purity	Immunogen affinity purified serum in PBS pH 7.4.
Format	Lyophilized
Quantity	50 µg
Reconstitution	For reconstitution add 50 µl of sterile water
Storage	Store lyophilized/reconstituted at -20°C; once reconstituted make aliquots to avoid repeated freeze-thaw cycles. Please remember to spin the tubes briefly prior to opening them to avoid any losses that might occur from material adhering to the cap or sides of the tube.
Additional information	This product can be sold containing ProClin if requested

Application information

Recommended dilution	1 : 3000 (WB)
Expected | apparent MW	44 kDa
Confirmed reactivity	*Arabidopsis thaliana*
Predicted reactivity	*Noccaea caerulescens* Species of your interest not listed? Contact us
Not reactive in	*Brachypodium distachyon, Nicotiana benthamiana, Solanum lycopersicum*
Additional information	Antibody is also recognizng phosphorylated form of BIK1
Selected references	González-Fuente et al. (2026). Bacteria use P-body condensates to attenuate host translation during infection. Sci Adv . 2026 Apr 24;12(17):eaec4477. doi: 10.1126/sciadv.aec4477. Zhang et al. (2026). The RING-Type Ubiquitin Ligase XBAT35. 2 Positively Modulates Plant Immunity by Promoting FLS2 Protein Accumulation. Mol Plant Microbe Interact . 2026 Mar 3. doi: 10.1094/MPMI-02-26-0015-R. Toth et al. (2026). Motif-based substrate mapping of the receptor-like cytoplasmic kinase BIK1 reveals novel components and regulatory nodes of plant immunity. Nat Plants. 2026 Feb 9. doi: 10.1038/s41477-025-02218-z. Salguero-Linares et al. (2025). Lack of AtMC1 catalytic activity triggers autoimmunity dependent on NLR stability. EMBO Rep . 2025 May;26(9):2378-2412. doi: 10.1038/s44319-025-00426-4. Ngou et al. (2021) Mutual potentiation of plant immunity by cell-surface and intracellular receptors. Nature. 2021 Mar 10. doi: 10.1038/s41586-021-03315-7. Epub ahead of print. PMID: 33692545. Wang et al. (2021) Arabidopsis PUB2 and PUB4 connect signaling components of pattern-triggered immunity. New Phytol. 2021 Dec 17. doi: 10.1111/nph.17922. Epub ahead of print. PMID: 34918346



Total protein from 7-day old *Arabidopsis thaliana* seedlings Col-0 (wild-type) and bik1 (null mutant) (Lu et al, 2009) were extracted with 50mM HEPES-KOH buffer containing 250 mM sucrose, 5% glycerol, 50 mM NaPP, 1 mM NaMo, 25 mM NaF, 10mM EDTA, 0.5% PVP, 3mM DTT, 1mM PMSF, 10uM Leupeptin & 10nM Calyculin, and then fractionated by ultracentrifugation at 100,000 x gravity for 30 min at 4°C into soluble (S100) and microsomal (P100) proteins as described in LaMontagne et al. (2016). 30 µg proteins of Total, S100, and P100 fractions were denatured at 65°C for 5 min, separated on a 10 % SDS-PAGE and blotted 1h to nitrocellulose using tank transfer. Blots were blocked with 1x PBS (from Fisher

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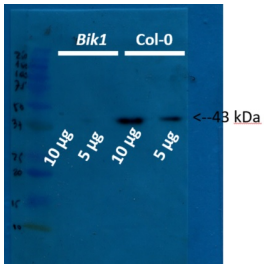
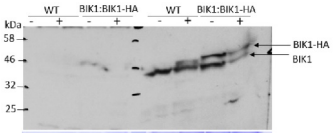
Scientific BP665-1) + 0.1 %Tween 20 (PBS-T) + 5% milk for 1h at room temperature (RT) with agitation. Blot was incubated in the primary antibody at a dilution of 1: 1000 overnight at 4°C with agitation in 1x PBS-T + 5% milk. The antibody solution was decanted, and the blot was rinsed briefly once, then washed three times for 7 min in 1x PBS-T at RT with agitation. Blot was incubated with secondary antibody (Goat anti Rabbit IgG (H&L) –HRP conjugated from Agrisera (AS09 602) diluted to 1:10 000 in 1x PBS-T + 5% milk for 2 hr at RT with agitation. The blot was washed as above and developed for 5 min with Amersham ECL (RPN2106). Exposure time was 6 min.

Courtesy of Gayani Ekanayake & Dr. Antje Heese Division of Biochemistry, Interdisciplinary Plant Group (IPG) - University of Missouri; Columbia, MO, USA



5 and 10 µg of protein from total plant extract of i*n vitro* grown *Arabidopsis thaliana* Col0 and bik1 mutant were extracted using basic phenol protocol. 1g were grinded with liquid nitrogen with mortair and pestle, powder were resuspended in 3 mL of protein buffer (0.5 M Tris-HCl, 0.7 M sucrose, 1 mM PMSF, 50 mM EDTA, 0.1 M KCl and 0.2% -mercaptoethanol; pH 8.0). Homogenate were centifuged at 8000 rpm and supernatant were mixed with three volumes of basic phenol, the mixture were incubated with agitation at room temperature during 10 mins. The mixture were centrifuged at 10000 rpm at 4ºC during 30 min, phenolic phase were recuperated in new tube and were mixed with one volume of ammonium acetate 0.1M in methanol, the mixture were incubated at -20 ºC during 4 hours. The mixture were centrifuged at 10000 rpm at 4º C during 20 minutes and protein pellet were washed three times with ammonium acetate 0.1M in methanol. Finally proteins were resuspended in tris-HCl (50 mM, pH 8.0) and the protein concentration were determined according Bradford method. Samples of 5 and 10µg were separated on 15 % SDS-PAGE using tank (BioRad system) to transfer to nitrocellulose membrane during 1 hour at 400 A. Blots were blocked with skimmed milk (10%) for 1h at room temperature (RT) with agitation. Blot was incubated in the primary antibody at a dilution of 1: 2 500 for 1h at RT with agitation. The antibody solution was decanted and the blot was rinsed briefly twice, then washed once for 15 min and 3 times for 5 min in TBS-T at RT with agitation. Blot was incubated in secondary antibody (anti-rabbit IgG horse radish peroxidase conjugated, from Agrisera AS09 602) diluted to 1:20 000 in for 1h at RT with agitation. The blot was washed as above and developed for 5 min with ECL according to the manufacturer's instructions. Exposure time was 20 seconds. For transference control the membrane was stained with Ponceau red and integrity of proteins was evaluated using 12% SDS-PAGE comassie blue stained.

Courtesy of Dr. Rodrigo A. Contreras, Laboratorio de Fisiología y Biotecnología Vegetal Departamento de Biología, Facultad de Química y Biología, Universidad de Santiago de Chile



Two-week-old seedlings of *Arabidopsis thaliana* (grown under long day conditions) were infiltrated for 10 mins with flg22 at 1uM or mock, collected and frozen in liquid nitrogen. Approximately 1g of plant material was used for microsomal extraction according to the protocol described by LaMontage et al., 2016). Samples (60ul) were separated on 16 % SDS-PAGE and blotted 2h to PVDF using tank transfer. Blots were blocked with for 1h at room temperature (RT) with agitation. Blot was incubated in the primary antibody at a dilution of 1: 3 000 at 4C ON with agitation in TBS-T + 1% BSA. The antibody solution was decanted and the blot was rinsed briefly twice, then washed once for 15 min and 3 times for 5 min in TBS-T at RT with agitation. Blot was incubated in secondary antibody (anti-rabbit IgG horse radish peroxidase conjugated, AS09 602, Agrisera) diluted to 1:20 000 in for 1h at RT with agitation. The blot was washed as above and developed for 2 mins with SuperSignal West Pico Chemiluminescent Substrate. Exposure time was ~ 1 hour.

Antibody is also recognizing phosphorylated form of BIK1. Applied treatment is decreasing pool of non-phosphorylated BIK1 and phosphoryled form is increasing.

Courtesy of Dr. Elwira Smakowska, Gregor Mendel Institute of Molecular Plant Biology , Vienna, Austria