

product **AS05 096**  
**TOP2 | DNA topoisomerase II**

## product information

<b>Background</b>	<b>Topoisomerase</b> type II (EC5.99.1.3) is one of the enzymes which is catalyzing unknotting of DNA by creating transient breaks in the DNA using a conserved tyrosine as the catalytic residue. Synonym names of this protein: At3g23890, ATTOPII, DNA topoisomerase 2, DNA topoisomerase II, F14O13.7, TOP2, TOPOISOMERASE II
<b>Immunogen</b>	The C-terminal 153 amino acids of the <i>Arabidopsis thaliana</i> Topoisomerase II (At3g23890, protein accession number <a href="#">P30182</a> ) with an N-terminal hexahistidine tag was expressed in <i>E.coli</i> and purified by Ni <sup>2+</sup> affinity chromatography.
<b>Host</b>	Rabbit
<b>Clonality</b>	Polyclonal
<b>Purity</b>	Serum
<b>Format</b>	Lyophilized
<b>Quantity</b>	200 µl
<b>Reconstitution</b>	For reconstitution add 200 µl of sterile water.
<b>Storage</b>	Store lyophilized/reconstituted at -20 °C; once reconstituted make aliquots to avoid repeated freeze-thaw cycles. Please, remember to spin tubes briefly prior to opening them to avoid any losses that might occur from lyophilized material adhering to the cap or sides of the tubes.
<b>Tested applications</b>	Immunolocalization (IL), Western blot (WB)
<b>Related products</b>	<a href="#">AS07 225</a>   anti-A12.2   RNA polymerase I subunit (homolog of Pol II Rpb9) rabbit antibody <a href="#">AS07 265</a>   anti-eEF1b   elongation factor eEF1b-beta protein rabbit antibody <a href="#">Plant protein extraction buffer</a> <a href="#">Secondary antibodies</a>
<b>Additional information</b>	Antibody detects a protein of ca. 170 kDa on western blots of <i>Arabidopsis thaliana</i> protein extracts. In subcellular fractions of cultured <i>Arabidopsis thaliana</i> cells the antibody detects a 170 kDa protein exclusively in the nuclear fraction (see picture).

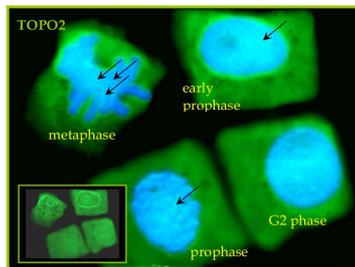
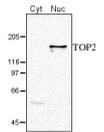
## Application information

<b>Recommended dilution</b>	1 : 500 (IL), 1 : 2000 (WB)
<b>Expected   apparent MW</b>	164   170 kDa ( <i>Arabidopsis thaliana</i> )
<b>Confirmed reactivity</b>	<i>Arabidopsis thaliana</i> , <i>Vicia faba</i>
<b>Predicted reactivity</b>	<i>Brassica rapa</i> , <i>Chlamydomonas reinhardtii</i> , <i>Chlorella vulgaris</i> , <i>Citrus clementina</i> , <i>Glycine max</i> , <i>Hordeum vulgare</i> , <i>Medicago truncatula</i> , <i>Oryza sativa</i> , <i>Ostreococcus tauri</i> , <i>Panicum italicum</i> , <i>Phaseolus vulgaris</i> , <i>Physcomitrella patens</i> , <i>Pinus sitchensis</i> , <i>Populus trichocarpa</i> , <i>Solanum tuberosum</i> , <i>Sorghum bicolor</i> , <i>Triticum aestivum</i> , <i>Vitis vinifera</i> , <i>Volvox Carterii</i>
<b>Not reactive in</b>	<i>Nicotiana tabacum</i>
<b>Additional information</b>	Topoisomerase II is highly expressed in young seedlings. The protein is localized in the nucleus and gene expression levels are increased in proliferative tissues like shoot apex or root tip. This product can be sold containing ProClin if requested.
<b>Selected references</b>	<a href="#">Xie S &amp; Lam E (1994)</a> Abundance of nuclear DNA topoisomerase II is correlated with proliferation in <i>Arabidopsis thaliana</i> . NAR 25:5729. <a href="#">Klaus Feldmann (1997)</a> Regulation der Topoisomerase II von <i>Arabidopsis thaliana</i> im

■ Zellzyklus. PhD thesis, University of Cologne.

## Application example

**10 µg of total protein** from (**Cyt**) *Arabidopsis thaliana* cytoplasmic fraction, (**Nuc**) *Arabidopsis thaliana* nuclear fraction were separated on SDS-PAGE and blotted 1h to **nitrocellulose**. Filters were probed with anti-TOP2 antibodies (AS05 096, **1:2000**). Signal was detected with SuperSignal West Pico ECL (Pierce).



Seeds of field bean (*Vicia faba* L. subsp. minor var. Nadwiślański; DANKO Group; Sobiejuchy) were sterilized using sodium hypochlorite (0.3% v/v) and germinated in Petri dishes on wetted filter paper at room temperature. At 4 d after imbibition, dark-grown seedlings with primary roots 25±5 mm long were selected for experiments. During incubations roots were oriented horizontally in a humid chamber and aerated continuously on a rotary water-bath shaker (30 rpm) at 23 °C. **Immunocytochemical assays** were performed according to the method prescribed earlier (Rybaczek and Maszewski 2006). Excised apical parts of roots (1.5 mm long) were fixed for 45 min (18 °C) in PBS-buffered 3.7% paraformaldehyde, washed several times with PBS and placed in a citric acid-buffered digestion solution (pH 5.0; 37 °C for 45 min) containing 2.5% pectinase (Fluka), 2.5% cellulase (Onozuka R-10; Serva) and 2.5% pectoliase (ICN). After removing the digestion solution, root tips were washed 3 times in PBS, rinsed with distilled water and squashed onto Super Frost Plus glass slides (Menzel-Gläser). Air-dried slides were pretreated with PBS-buffered 5% BSA at 20 °C for 50 min and incubated overnight in a humidified atmosphere (4 °C) with rabbit antibody raised against TOPO2 (Agrisera), dissolved in PBS containing 1% BSA (at a dilution of 1:500). Following incubation, slides were washed 3 times with PBS and incubated for 1 h (18 °C) with secondary goat anti-rabbit IgG DyLight®488 antibody (Agrisera, [AS09 633](#), 1:3000). Nuclear DNA was stained with 4',6-diamidino-2-phenyl-indole (DAPI, 0.4 µg/ml; Sigma-Aldrich). Following washing with PBS, slides were air dried and embedded in Vectashield Mounting Media for Fluorescence (Vector Laboratories). Observations were made using Optiphot-2 fluorescence microscope (Nikon) equipped with B-2A filter (blue light; 495 nm) for DyLight-conjugated antibodies and UV-2A filter (UV light; 365 nm) for DAPI. All images were recorded at exactly the same time of integration using DXM 1200 CCD camera.

Courtesy Dr. Dorota Rybaczek, Lodz University, Poland