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### Product no AS07 204

# Anti-FtsH3 + FtsH10 | ATP-dependent zinc metalloprotease FtsH3 + FtsH10 (mitochondrial)

#### **Product information**

Immunogen KLH-conjugated peptide dereived from sequences of Arabidopsis thaliana FtsH3 and FtsH10 with localization to mitochondria Q84WU8, At2g29080 and Q8VZI8, At1g07510

**Host** Rabbit

Clonality Polyclonal

**Purity** Immunogen affinity purified serum in PBS pH 7.4.

Format Lyophilized

Quantity 200 μg

Reconstitution For reconstitution add 100 µl of sterile water

Store lyophilized/reconstituted at -20°C; once reconstituted make aliquots to avoid repeated freeze-thaw cycles. Please Storage remember to spin the tubes briefly prior to opening them to avoid any losses that might occur from material adhering to

the cap or sides of the tube.

Additional information Blue-native (2D BN/SDS-PAGE) methodology is described in Piechota et al. 2010

## **Application information**

Recommended dilution 1:500-1:1000 (WB)

Expected | apparent 80 kDa

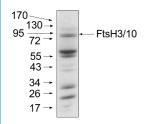
Not reactive in No confirmed exceptions from predicted reactivity are currently known

Selected references Kolodziejczak et al. (2018). m-AAA Complexes Are Not Crucial for the Survival of Arabidopsis Under Optimal Growth

Conditions Despite Their Importance for Mitochondrial Translation. Plant Cell Physiol. 2018 May 1;59(5):1006-1016. doi: 10.1093/pcp/pcy041.

Piechota et al. (2010). Identification and characetization of high-molecular-weight complexes fromed by m-AAA proteases and prohibitins in mitochondria of Arabidopsis thaliana. J Biol Chem. 2010 Apr 23;285(17):12512-21. doi:

## **Application example**



Total protein from Arabidopsis thaliana mitochondria (20 µg) were separated on 10% acrilamide gel and electrophoresis prepared according to Schägger and von Jagov (Anl. Biochem., 1987, 166:368-379). After running the gel, proteins were transferred to nitrocellulose membrane using wet transfer (0.22% CAPS, pH 11). Transfer was checked by Ponceau S staining. Blot was destained by several quick washings in distilled water and 1 washing in 1X TBS (10 mM T pH 7.5, 150 mM NaCl) (10-15 min.). Blot was blocked by 1.5 hour in 5% milk in TBST (1X TBS, 0,1 20) After blocking blot was washed quickly twice in TBST and incubated 2 hours with primary antibody (dilution 1: 1000 TBST (dilution 1:1000). Washing: two quick washings in TBST and 3 x 10 min. washings in TBST. Then blot was incubated 45-60 min. with a secondary anti-rabbit antibodies conjugated to peroxidase (dilution 1:10000) in TBST. Washing: as above. After washing blot was incubated 1-2 min. in ECL solution and exposed to Kodak autoradiography film. Exposure time was 15-60 seconds.

Mitochondria were isolated as described by Urantowka et al. (Plant Mol Biol, 2005, 59:239-52). Mitochondrial pellets were suspended in 1X Laemmli buffer (5% beta-mercaptoetanol, 3.7% glycerol, 1.1% SDS, 23 mM Tris-HCl pH 6.8, 0.01% bromophenol blue), heated (95°C, 5 min.) and centrifuged (13 000rpm, 1 min.).



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