

Product no **AS03 037****Anti-RbcL | Rubisco large subunit, form I (rabbit)****Product information**

<b>Immunogen</b>	KLH-conjugated synthetic peptide conserved across all known plant, algal and cyanobacterial RbcL protein sequences (form I L8S8 and form II L2), including, <i>Arabidopsis thaliana</i> <a href="#">Q03042</a> , <i>Hordeum vulgare</i> <a href="#">P05698</a> , <i>Oryza sativa</i> <a href="#">P0C510</a> , <i>Chlamydomonas reinhardtii</i> <a href="#">P00877</a> , <i>Synechococcus</i> PCC 7920 <a href="#">A5CKC5</a>
<b>Host</b>	Rabbit
<b>Clonality</b>	Polyclonal
<b>Purity</b>	Serum
<b>Format</b>	Lyophilized
<b>Quantity</b>	50 µl
<b>Reconstitution</b>	For reconstitution add 50 µl of sterile water
<b>Storage</b>	Store lyophilized/reconstituted at -20°C; once reconstituted make aliquots to avoid repeated freeze-thaw cycles. Please remember to spin the tubes briefly prior to opening them to avoid any losses that might occur from material adhering to the cap or sides of the tube.
<b>Additional information</b>	<p>Anti-RbcL can be used as a cellular [compartment marker] of plastid stroma (cytoplasm in cyanobacteria) and detects RbcL protein from 31.25 fmoles. As both forms (I and II) are detected it is suitable for work with samples from Dinoflagellates, Haptophytes and Ochrophytes (diatoms, Raphidophytes, brown algae) as well as higher plants. This antibody together with Agrisera Rubisco protein standard is very suitable to quantify Rubisco in plant and algal samples. Example of a simultaneous western blot detection with RbcL, PsbA and Psac antibodies.</p> <p>This antibody is not suitable for use in immunoprecipitation.</p> <p>This product can be sold containing ProClin if requested.</p>

**Application information**

<b>Recommended dilution</b>	Immunofluorescence/confocal microscopy (IF), 1: 1000 (IG), 1: 250 for images see <a href="#">Prins et al. (2008)</a> , detailed protocol available on request, 1: 800 (TP), 1: 5000 - 10 000 and more depending upon protein load and detection method (WB)
<b>Expected   apparent MW</b>	52.7 kDa ( <i>Arabidopsis thaliana</i> ), 52.5 kDa (cyanobacteria), 52.3 ( <i>Chlamydomonas reinhardtii</i> )
<b>Confirmed reactivity</b>	<i>Agostis stolonifera</i> cv. <i>Penncross</i> , <i>Arabidopsis thaliana</i> , <i>Apium graveolens</i> , <i>Artemisia annua</i> , <i>Atrichum undulatum</i> , <i>Attheya longicornis</i> , <i>Baculogypsina sphaerulata</i> (benthic foraminifer), <i>Beta vulgaris</i> , <i>Begonia</i> sp., <i>Bienertia sinuspersici</i> , <i>Brassica napus</i> , <i>Kandelia candel</i> , <i>Cannabis sativa</i> L., <i>Chaetoceros furcellatus</i> , <i>Chlorococcum dorsiventrale</i> , <i>Colobanthus quitensis</i> , <i>Cicer arietinum</i> , <i>Chenopodium quinoa</i> , <i>Chlamydomonas raudensis</i> , <i>Chlamydomonas reinhardtii</i> , <i>Colobanthus quitensis</i> Kunt Bartl, <i>Chlorella sorokiniana</i> , <i>Chlorella vulgaris</i> , <i>Coscinodiscus concinnus</i> , <i>Cyanophora paradoxa</i> , <i>Cylindrospermopsis raciborskii</i> CS-505, <i>Cynara cardunculus</i> , <i>Emiliana huxleyi</i> , <i>Euglena gracilis</i> , <i>Ficus carica</i> , <i>Fortunella margarita</i> Swingle, <i>Fraxinus mandshurica</i> , <i>Fucus vesiculosus</i> , <i>Gladieria sulphuraria</i> , <i>Glycine max</i> , <i>Gonyaulax polyedra</i> , <i>Gongolaria barbata</i> , <i>Guzmania</i> hybrid, <i>Heterosigma akashiwo</i> , <i>Hevea</i> , <i>Hordeum vulgare</i> , <i>Hypnum cupressiforme</i> , <i>Jatropha curcas</i> , <i>Karenia brevis</i> (C.C.Davis) s) G.Hansen & Ø.Moestrup (Wilson isolate), <i>Kochia prostrata</i> , <i>Lathyrus sativus</i> , <i>Liquidambar formosana</i> , <i>Malus domestica</i> , <i>Medicago truncatula</i> , <i>Micromonas pusilla</i> , <i>Nicotiana benthamiana</i> , <i>Nicotiana tabacum</i> , <i>Panicum virgatum</i> , <i>Petunia hybrida</i> cv. Mitchell, <i>Phaeodactylum tricorutum</i> , <i>Physcomitrium patens</i> , <i>Pisum sativum</i> , <i>olytrichum formosum</i> , <i>Porosira glacialis</i> , <i>Porphyra</i> sp., <i>Ricinus communis</i> , <i>Robinia pseudoacacia</i> , <i>Rhytidadelphus squarrosus</i> , <i>Saccharum</i> sp., <i>Schima superba</i> , <i>Skeletonema costatum</i> (diatom), <i>Skeletonema marinoi</i> (diatom), <i>Solanum lycopersicum</i> , <i>Spinacia oleracea</i> , lichens, <i>Stanleya pinnata</i> , <i>Symbiodinium</i> sp., <i>Synechococcus</i> PCC 7942, <i>Synechococcus elongatus</i> UTEX 2973, <i>Rhoeo discolor</i> , <i>Thalassiosira pseudonana</i> , <i>Thermosynechococcus elongatus</i> , <i>Triticum aestivum</i> , <i>Prochlorococcus</i> sp. (surface and deep water ecotype), <i>Triticum aestivum</i> , dinoflagellate endosymbionts (genus <i>Symbiodinium</i> ), extreme acidophilic verrucomicrobial methanotroph <i>Methylacidiphilum fumarolicum</i> strain SolV, <i>Thalassiosira punctigera</i> , <i>Tisochrysis lutea</i> , <i>Verbascum lychnitis</i> , <i>Vitis vinifera</i> , <i>Quercus ilex</i>
<b>Predicted reactivity</b>	Alpha proteobacteria, Algae (brown and red) including <i>Galdieria sulphuraria</i> , <i>Arthrospira platensis</i> , Dicots, <i>Benincasa hispida</i> , <i>Kalanchoe fedtschenkoi</i> ; Beta-proteobacteria, Conifers, Cryptomonads, Cyanobacteria (prochlorophytes), <i>Eragrostis tef</i> , Gamma-proteobacteria, Liverworts, <i>Manihot esculenta</i> , <i>Marchantia polymorpha</i> , <i>Miscanthus giganteus</i> , Monocots, Mosses, <i>Suaeda glauca</i> , <i>Welwitschia</i> ; <i>Nannochloropsis</i> sp., <i>Picochlorum</i> sp., <i>Porphyridium purpureum</i> , <i>Zea mays</i> , <i>Zosteria marina</i>
	For detection in Rhodospirillaceae use product <a href="#">AS15 2955</a>
	Species of your interest not listed? <a href="#">Contact us</a>

**Not reactive in** | No confirmed exceptions from predicted reactivity are currently known

**Additional information** | This antibody was used in:

Immunocytochemical staining of diatoms according to Schmid (2003) J Phycol 39: 139-153 and Wordemann et al. (1986) J Cell Biol 102: 1688-1698.

Immunofluorescence [Dreier](#) et al. (2012). FEMS Microbial Ecol., March 2012.

Western blot and tissue printing during a student course [Ma](#) et al. (2009).

As a loading control [Sun](#) et al. (2020).

Protocol for Rubisco quantification using this antibody can be found [here](#).

### Selected references

[Xiao](#) et al. (2025). A fungal effector promotes infection via stabilizing a negative regulatory factor of chloroplast immunity. Nat Commun. 2025 Jul 29;16(1):6970. doi: 10.1038/s41467-025-62326-4.

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[Krupinska](#) et al. (2025). Iron allocation to chloroplast proteins depends on the DNA-binding protein WHIRLY1. Planta. 2025 Jun 17;262(2):32. doi: 10.1007/s00425-025-04736-8.

[Archer](#) et al. (2025). Effects of chaperone selectivity on the assembly of plant Rubisco orthologs in E. coli. J Exp Bot . 2025 Mar 31:eraf140. doi: 10.1093/jxb/eraf140.

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[Pica](#) et al. (2024). Functional ecological traits in young and adult thalli of canopy-forming brown macroalga *Gongolaria barbata* (Phaeophyta) from a transitional water system. PeerJ. 2024 Sep 12:12:e17959. doi: 10.7717/peerj.17959.

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[Nuamzanei](#) et al. (2024). Impact of polyvinyl chloride (PVC) microplastic on growth, photosynthesis and nutrient uptake of *Solanum lycopersicum* L. (Tomato). Environ Pollut. 2024 Apr 16:123994. doi: 10.1016/j.envpol.2024.123994.

[Ciesielska](#) et al. (2024). S2P2-the chloroplast-located intramembrane protease and its impact on the stoichiometry and functioning of the photosynthetic apparatus of *A. thaliana*. Front Plant Sci. 2024 Mar 15:15:1372318. doi: 10.3389/fpls.2024.1372318.

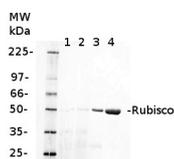
[Rodrigues](#) et al. (2023). Are tomato plants co-exposed to heat and salinity able to ensure a proper carbon metabolism?—An insight into the photosynthetic hub. Plant Physiol Biochem. 2023 Dec 10:206:108270. doi: 10.1016/j.plaphy.2023.108270.

[Redhi](#) et al. (2023). The UV-A Receptor CRY-DASH1 Up- and Downregulates Proteins Involved in Different Plastidial Pathways. J Mol Biol. 2023 Sep 10:168271. doi: 10.1016/j.jmb.2023.168271.

### Western blot



**0.25 µg of chlorophyll a/lane** from *Spinacia oleracea* (1), *Synechococcus* PCC 7942 (2), *Cyanophora paradoxa* (3), *Heterosigma akashiwo* (4), *Thalassiosira pseudonana* (5), *Euglena gracilis* (6), *Micromonas pusilla* (7), *Chlamydomonas reinhardtii* (8), *Porphyra* sp (9), *Gonyaulax polyedra* (10), *Emiliana huxleyi* (11) extracted with PEB (**AS08 300**), were separated on **4-12%** NuPage (Invitrogen) **LDS-PAGE** and blotted 1h to **nitrocellulose**. Filters were blocked 1h with 2% low-fat **milk powder** in TBS-T (0.1% TWEEN 20) and probed with anti-RbcL antibody (AS03 037, **1:50 000**, 1h) and secondary anti-rabbit (**1:20000**, 1 h) antibody (HRP conjugated, recommended secondary antibody **AS09 602**) in TBS-T containing 2% low fat milk powder. Antibody incubations were followed by washings in TBS-T. All steps were performed at RT with agitation. Blots were developed for 5 min with ECL Advance detection reagent according the manufacturers instructions (GE Healthcare). Images of the blots were obtained using a CCD imager (FluorSMax, Bio-Rad) and Quantity One software (Bio-Rad).



This product is **for research use only** (not for diagnostic or therapeutic use)

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1 µg of chlorophyll from Cryptophyte samples (**1,2**) and 1 µg of chlorophyll (**3**) or 10 µg of total protein (**4**) from *Arabidopsis thaliana* leaves extracted either with 2 ml of 100 mM TrisHCl, 50 mM EDTA, 250 mM NaCl, 0.05% SDS (Sample 1) or 10 mL of 50 mM Hepes-KOH (pH 7.8), 330 mM sorbitol, 10 mM EDTA, 5 mM NaCl, 5 mM MgCl<sub>2</sub>, 5 mM sodium ascorbate and 0.2% BSA (Sample 2). Samples were denatured with 1:1 Amersham WB Loading Buffer at 70°C for 10 min and were separated on pre-casted 13.5% Amersham WB gel and blotted for 30 min to Amersham WB PVDF using wet transfer. Blots were blocked with 2% Amersham ECL Blocking Agent for 1h at room temperature (RT) with agitation. Blot was incubated in the primary antibody at a dilution of 1: 10 000 (rabbit anti-Rubisco AS03 037) for 1.5 h at RT with agitation. The antibody solution was decanted and the blot was rinsed briefly twice, then washed once for 15 min and 3 times for 5 min in TBS-T at RT with agitation. Membrane was cut in half and left part was incubated in anti-rabbit DyLight® 550 secondary antibody from Agrisera ([AS11 1782](#)) diluted to 1:2 000 in TBST for 1h at RT with agitation. The blot was scanned using Cy3 channel of Amersham WB System.

Courtesy of Dr. Małgorzata Wessels, Agrisera



**2 µg of total protein** from various plant extracts (**1-5**) extracted with PEB ([AS08 300](#)) separated on **4-12% NuPage** (Invitrogen) **LDS-PAGE** and blotted 1h to **PVDF**. Markers MagicMarks (Invitrogen) (**M**) and Rubisco protein standard (AS01 017S) at **0.0625 pmol, 0.125 pmol, 0.25 pmol**.

Following standard western blot procedure this image has been obtained using a CCD imager (FluorSMax, Bio-Rad) and Quantity One software (Bio-Rad). The contour tool of the software is used to the area for quantitation and the values are background subtracted to give an adjusted volume in counts for each standard and sample.

**Note:** Optimal quantitation is achieved using moderate sample loads per gel lane, generally 0.5 to 2.5 µg total protein, depending on the abundance of the target protein.