

This product is for research use only (not for diagnostic or therapeutic use)

contact: support@agrisera.com

Agrisera AB | Box 57 | SE-91121 Vännäs | Sweden | +46 (0)935 33 000 | www.agrisera.com

Product no AS23 4914

Anti-PFK1-7 | Phosphofructokinase 1-7

Product information

Immunogen KLH-conjugated peptide derived from Arabidopsis thaliana PFK1-7 protein sequences, UniProt: Q9M0F9,

Q9FIK0.Q94AA4, Q9FKG3 Q8VYN6 Q9M076 Q9C5J7 TAIR: At4g29220, At5g47810, At4g26270, At5g61580,

At2g22480, At4g32840, At5g56630

Host Rabbit

Clonality Polyclonal

Purity Antigen affinity purified serum, in PBS pH 7.4

Format Lyophilized

Quantity 50 μg

Reconstitution For reconstitution add 50 μl, of sterile or deionized water.

Storage Store lyophilized/reconstituted at -20°C; once reconstituted make aliquots to avoid repeated freeze-thaw cycles. Please, remember to spin tubes briefly prior to opening them to avoid any losses that might occur from lyophilized

material adhering to the cap or sides of the tubes.

Application information

Recommended dilution 1:1000 - 1:2000 (WB)

Expected | apparent

51.9 kDa

паріобры і папапа

Predicted reactivity Brassica napus (PFK1, PFK3, PFK5, PFK6, PFK7),

Nicotiana tabacum (PFK3, PFK4, PFK5, PFK6) Pisum sativum (PFK3, PFK4, PFK5, PFK7, PFK6)

Solanum lycopersicum (PFK3, PFK4m PFK5, PFK6 Solanum tuberosum (PFK3, PFK4, PFK5, PFK6)

Arachis hypogaea, Brachypodium distachyon, Brassica napus, Cannabis sativa, Hordeum vulgare, Malus domestica, Manihot esculenta, Medicago truncatula, Nicotiana tabacu, Oryza sativa, Saccharum sp., Theobroma cacao,

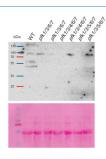
Triticum sp, ,Sorghum bicolor, Zea mays, Vitis vinifera

Species of your interest not listed? Contact us

Not reactive in No confirmed exceptions from predicted reactivity are currently known

Selected references To be added when available, antibody available in January 2025.





Samples:

20 ug of Arabidopsis thaliana whole leaf extract

20 ug of Arabidopsis thaliana mutant described above the picture MW markers are marked on the left side of each membrane

20 μg/well of total protein extracted freshly from leaf extracts of *Arabidopsis thaliana* wildtype and mutants. Exact buffer components were: 50 mM Hepes-KOH (pH 6.8), 5 mM Mg-acetate, 15 % Glycerin, 1 mM EDTA, 1 mM EGTA, 5 mM β-Mercaptoethanol, 0.1 mM Pefabloc Proteinase-inhibitor and denatured with 1 x Laemmli-buffer (62.5 mM Tris-HCl (pH 6.8), 2 % SDS, 10 % Glycerin, 5 % β-Mercaptoethanol, 0.001 % Bromphenolblue) at 98 °C / 2 mins. Samples were separated on 10 % SDS-PAGE and blotted for 1 h PVDF (pore size of 0.45 μm), using: wet transfer in the cold. Blot was blocked with 5 % nonfat milk 4 °C / ON with agitation. Blot was incubated in the primary antibody at a dilution of 1: 2 000 for 1 h/RT with agitation in TBS-T. The antibody solution was decante, andd the blot was rinsed briefly, then washed once 3 times for 10 min in TBS-T at RT with agitation. Blot was incubated in matching secondary antibody (anti-rabbit IgG horse radish peroxidase conjugated) diluted to 1: 25 000 in for 1 h/RT with agitation. The blot was washed as above and developed with a following chemiluminescent detection reagent.



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Exposure time was 3 minutes.

Courtesy of phd student Alina Johanna Hieber, University of Bayreuth, Bayreuth, Germany