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Product no AS22 4713

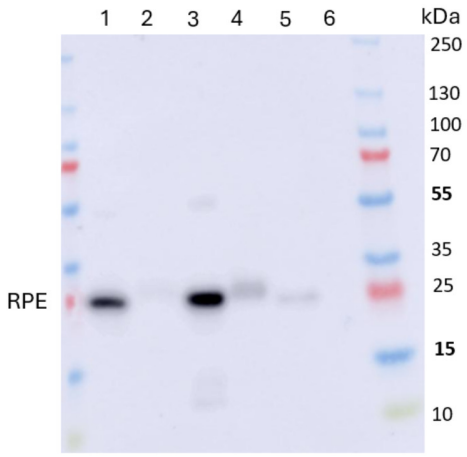
Anti-RPE | Ribulose-5-phosphate-3-epimerase, chloroplastic

Product information

Immunogen	KLH-conjugated peptide derived from *Arabidopsis thaliana* RPE protein sequence, UniProt: Q9SAU2, TAIR: At5g61410
Host	Rabbit
Clonality	Polyclonal
Purity	Affinity purified serum, in PBS pH 7.4
Format	Lyophilized
Quantity	50 µg
Reconstitution	For reconstitution, add 50 µl of sterile or deionized water.
Storage	Store lyophilized/reconstituted at -20°C; once reconstituted make aliquots to avoid repeated freeze-thaw cycles. Please, remember to spin tubes briefly prior to opening them to avoid any losses that might occur from lyophilized material adhering to the cap or sides of the tubes.

Application information

Recommended dilution	1: 500 - 1 : 1000 (WB)
Expected | apparent MW	30 | 25 kDa (due to N-terminal or C-terminal processing)
Confirmed reactivity	*Arabidopsis thaliana, Nicotiana benthamiana*
Predicted reactivity	*Capsicum annuum, Chlamydomonas reinhardii, Cucumis sativus, Lolium perenne, Physcomitrium patens, Solanum lycopersicum, Solanum tuberosum, Spinacia oleracea* Species of your interest not listed? Contact us
Not reactive in	*Nannochloropsis* sp., *Vicia faba*
Selected references	To be added when available, antibody available in October 2025.



Samples:
1 - 10 µg of *Spinacia oleracea* total cell extract
2 - 10 µg of *Solanum lycopersicum* total cell extract
3 - 10 µg of *Cucumis sativus* total cell extract
4 - 10 µg of *Solanum tuberosum* total cell extract
5 - 10 µg of Rubisco protein standard (SIGMA)
6 - 10 µg of BSA (negative control)

10 µg/well of total protein extracted from frozen leaves from *Spinacia oleracea, Solanum lycopersicum, Cucumis sativus, Solanum tuberosum*. Leaves were pressed into a juice using an Angel Juicer (5500). Samples were normalised to 1mg/mL based on a Bradford assay, using ddH20 (pH 7.5) as dilutant. Samples were mixed in a 3:1 (v/v) ratio with Laemmli loading buffer containing 8% SDS, 1% bromophenol blue, 10 mM EDTA, 250 mM Tris-HCl (pH 6.8), 40% glycerol, and 20% -mercaptoethanol. Mixtures were heated at 95 °C for 5 minutes, briefly centrifuged (4000xg, 5mins) and loaded onto 15-well 4–20% Mini-PROTEAN® TGX™ Precast Protein Gels (Bio-Rad, 4561096) using a Bio-Rad Mini-Protein

cell (Bio-Rad, Hercules, USA). Protein transfer was carried out on nitrocellulose (pore size of 0.45 µm - Nitrocellulose Membrane, Roll (Bio-Rad) (#1620115) & Cytiva Whatman™ Grade GB003 Blotting Paper (11959257)), using: semi-dry or dry transfer (TGX Turbo Setting). Blot was blocked with 5 % milk (in 1x PBS) 1h/RT with agitation. Blot was incubated in the primary antibody at a dilution of 1:500 for 1h/RT with agitation in 5% milk (in 1x PBS). The antibody solution was decanted, and the blot was rinsed briefly twice, then washed once for 15 min and 3 times for 5 min in PBS-T at RT with agitation. Blot was incubated in matching secondary antibody (anti-rabbit IgG horse radish peroxidase conjugated) diluted to 1:25000 in 5% milk (in 1x PBS) for 1h/RT with agitation. The blot was washed as above and developed with a following chemiluminescent detection reagent: SuperSignal™ West Pico PLUS Chemiluminescent Substrate. Exposure on the Image Quant LAS 500 instrument using the 'Auto' chemiluminescence option.

Note: For detection of RPE in different species, protein load/well and primary antibody dilution and incubation time must be optimized.

Courtesy of Jack Pattinson, IAFRI PhD Student, Molecular Life Sciences (MLS), School of Natural and Environmental Sciences Devonshire Building, Newcastle University, United Kingdom