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contact: support@agrisera.com

Agrisera AB | Box 57 | SE-91121 Vännäs | Sweden | +46 (0)935 33 000 | www.agrisera.com

Product no AS08 325

## Anti-ARF1 | ADP-ribosylation factor 1

### **Product information**

Immunogen Recombinant GST fusion of full length of Arabidopsis thaliana ARF1 (P36397, AT2G47170)

**Host** Rabbit

Clonality Polyclonal

**Purity** Serum

Format Lyophilized

Quantity 50 μl

**Reconstitution** For reconstitution add 50 μl of sterile water

Storage Store lyophilized/reconstituted at -20°C; once reconstituted make aliquots to avoid repeated freeze-thaw cycles. Please remember to spin the tubes briefly prior to opening them to avoid any losses that might occur from material adhering to

the cap or sides of the tube

Additional information Cellular [compartment marker] of Golgi in immunolocalization and COP1 in western blot

### Application information

**Recommended dilution** 1:1000 (IF), 1:100 (IG), 1:1000 (WB)

Expected | apparent

21 kDa (Arabidopsis thaliana)

**Confirmed reactivity** Actinidia chinensis, Arabidopsis thaliana, Chlamydomonas reinhardtii, Elaeis sp., Lilium longiflorum, Medicago

truncatula, Nicotiana tabacum, Oryza sativa, Petunia hybrida cv Mitchell, Physcomitrium patens, Solanum tuberosum

Brassica juncea, Brassica napus, Capsella rubella, Capsicum annum, Cucumis sp., Daucus carota, Elaeis guineensis, Predicted reactivity Glycine max, Helleborus orientalis, Hordeum vulgare, Medicago truncatula, Nannochloropsis gaditana, Nicotiana

benthamina, Ostreococcus tauri, Populus trichocarpa, Triticum aestivum, Zea mays

Species of your interest not listed? Contact us

Not reactive in Centella asiatica, Microsporidia sp.

**Additional information** References describing immunolocalization (IF) and (IG) studies:

> Pimpl et al (2000). In Situ Localization and in Vitro Induction of Plant COPI-Coated Vesicles. Plant Cell. 2000 Nov:12(11):2219-36.

Ritzenthaler et al. (2002). Reevaluation of the Effects of Brefeldin A on Plant Cells Using Tobacco Bright Yellow 2 Cells

Expressing Golgi-Targeted Green Fluorescent Protein and COPI Antisera. Plant Cell. 2002 Jan;14(1):237-61.

Selected references

Mohr et al. (2024). Characterization of the small Arabidopsis thaliana GTPase and ADP-ribosylation factor-like 2 protein TITAN. J Cell Sci. 2024 Aug 14;137(15):jcs262315. doi: 10.1242/jcs.262315. (immunofluorescence) Suanno et al. (2023) Small extracellular vesicles released from germinated kiwi pollen (pollensomes) present characteristics similar to mammalian exosomes and carry a plant homolog of ALIX, Front. Plant Sci., 25 January 2023, Sec. Plant Membrane Traffic and Transport, Volume 14 - 2023.

Chien et al. (2022) Phosphate transporter PHT1;1 is a key determinant of phosphorus acquisition in Arabidopsis natural accessions. Plant Physiol. 2022 Aug 29;190(1):682-697. doi: 10.1093/plphys/kiac250. PMID: 35639954; PMCID: PMC9434223. (Immunogold).

Brumm, Singh, Kriechbaum, et al. (2022) N-terminal domain of ARF-GEF GNOM prevents heterodimerization with functionally divergent GNL1 in Arabidopsis. Plant J. 2022;112(3):772-785. doi:10.1111/tpj.15979.

Farago et al. (2022) Small paraquat resistance proteins modulate paraquat and ABA responses and confer drought tolerance to overexpressing Arabidopsis plants. Plant Cell Environ. 2022 Jul;45(7):1985-2003. doi: 10.1111/pce.14338. Epub 2022 Apr 29. PMID: 35486392; PMCID: PMC9324991.

Narasimhan et al. (2021) Systematic analysis of specific and nonspecific auxin effects on endocytosis and trafficking. Plant Physiol. 2021 Mar 18:kiab134. doi: 10.1093/plphys/kiab134. Epub ahead of print. PMID: 33734402.

Hurny et al. (2020). SYNERGISTIC ON AUXIN AND CYTOKININ 1 Positively Regulates Growth and Attenuates Soil Pathogen Resistance. Nat Commun. 2020 May 1;11(1):2170. doi: 10.1038/s41467-020-15895-5. (immunolocalization) Kuang et al. (2019). Quantitative Proteome Analysis Reveals Changes in the Protein Landscape During Grape Berry Development With a Focus on Vacuolar Transport Proteins. Front Plant Sci. 2019 May 15;10:641. doi: 10.3389/fpls.2019.00641. eCollection 2019.

Singh et al. (2018). A single class of ARF GTPase activated by several pathway-specific ARF-GEFs regulates essential membrane traffic in Arabidopsis. PLoS Genet. 2018 Nov 15;14(11):e1007795. doi: 10.1371/journal.pgen.1007795. Lang et al. (2011). Simultaneous isolation of pure and intact chloroplasts and mitochondria from moss as the basis for



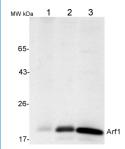
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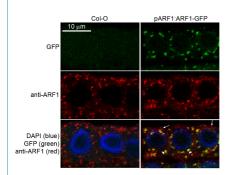
sub-cellular proteomics. Plant Cell Rep. 2011 Feb;30(2):205-15.doi: 10.1007/s00299-010-0935-4.

# **Application example**



50 μg of total protein from (1) *Nicotiana tabacum* protoplast total protein, (2) *Arabidopsis thalian*a protoplast soluble protein, (3) *Arabidopsis thaliana* protoplast total protein were separated on 10 % SDS-PAGE and blotted 2h to nitrocellulose (Semi-dry, 200mA). Filters were blocked over night with 5% low-fat milk powder in TBS and probed with anti-Arf1 antibodies (AS08 325, 1:1000, 1h) and secondary anti-rabbit (1:20000, 1h) antibody (HRP) in TBS-Tween (recommended secondary antibody AS09 602). Signal was detected with chemiluminescence detection reagent and exposure time for this image was 1 minute.

Protoplasts were extracted in 50mM Tris, 10 mM EDTA and Triton X100, 0.02%.



### Immunofluorescence

Specificity testing of rabbit anti-ARF1 serum. Immunofluorescence labelling of rabbit anti-ARF1 antibody (red) in 5-day-old root epidermal cells of the *Arabidopsis thaliana* ecotype Columbia-0 (WT) or seedlings expressing the ADP-RIBOSYLATION FACTOR 1 (AtARFA1c; accession At2g47170) fused to EGFP (green) (Xu, J. and Scheres, B. 2005. Plant Cell 17, 525-536). The rabbit anti-ARF1 antibody was diluted 1:1000 and the secondary antibody, donkey anti-rabbit CY5-coupled (Jackson ImmunoResearch) was diluted 1:300. The nuclei were stained with DAPI (blue). Note the co-labelling of ARF1-GFP with the anti-ARF1 antibody (arrowheads) and the additional labelling (potentially of other ARF1 variants) by the anti-ARF1 antibody (arrows). The antibody staining permeability was limited to the 1-2 outermost layers of the whole-mounted root tips.

Courtesy of Dr. Anna Gustavsson and Dr. Markus Grebe