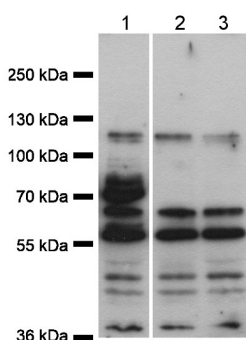


Product no **AS19 4315****CPK1 | Calcium-dependent protein kinase 1****Product information**

<b>Immunogen</b>	KLH-conjugated peptide derived from <i>Arabidopsis thaliana</i> CPK1 protein sequence, UniProt: <a href="#">Q06850-1</a> , TAIR <a href="#">AT5G04870</a>
<b>Host</b>	Rabbit
<b>Clonality</b>	Polyclonal
<b>Purity</b>	Immunogen affinity purified serum in PBS pH 7.4.
<b>Format</b>	Lyophilized
<b>Quantity</b>	50 µg
<b>Reconstitution</b>	For reconstitution add 50 µl, of sterile water
<b>Storage</b>	Store lyophilized/reconstituted at -20°C; once reconstituted make aliquots to avoid repeated freeze-thaw cycles. Please remember to spin the tubes briefly prior to opening them to avoid any losses that might occur from material adhering to the cap or sides of the tube.

**Application information**

<b>Recommended dilution</b>	1 : 1000 (WB)
<b>Expected   apparent MW</b>	68   72 kDa
<b>Confirmed reactivity</b>	<i>Arabidopsis thaliana</i>
<b>Predicted reactivity</b>	<i>Noccaea caerulescen</i> , <i>Triticum</i> sp. Species of your interest not listed? <a href="#">Contact us</a>
<b>Not reactive in</b>	No confirmed exceptions from predicted reactivity are currently known
<b>Selected references</b>	<a href="#">Durian et al. (2019)</a> . PROTEIN PHOSPHATASE 2A-B'Ä?Ä³ controls Botrytis cinerea resistance and developmental leaf senescence. <i>Plant Physiol.</i> 2019 Oct 28. pii: pp.00893.2019. doi: 10.1104/pp.19.00893. <a href="#">Durian et al. (2019)</a> . PROTEIN PHOSPHATASE 2A-B'Ä?Ä³ controls Botrytis cinerea resistance and developmental leaf senescence. <i>Plant Physiol.</i> 2019 Oct 28. pii: pp.00893.2019. doi: 10.1104/pp.19.00893.

**Application example**

Plant age: 4 weeks under short-day conditions (8 h light).

20 µg of the total protein from *Arabidopsis thaliana* plants, (genetic background: Col-0) plants, wild-type (1), *cpk1* (SALK\_096452) (2), *cpk1* (SALK\_080155c) (3) were extracted with protein extraction buffer (Durian et al., 2019) and denatured in Laemmli-sample buffer (final concentrations: 5% (v/v) beta-mercaptoethanol, 69 mM Tris-HCl pH 6.8, 11.1% (v/v) glycerol, 2.15% (v/v) SDS) for 10 min at 74 °C. Samples were separated by discontinuous gel electrophoresis on a 10% (w/v) separation SDS-PA-gel and blotted 1h to PVDF-membrane using a semi-dry system. The blot was blocked with 5% milk powder in TBS-T with agitation for 1h at room temperature (RT). After rinsing in TBS-T, the blot was incubated in the primary antibody at a dilution of 1:1000 in TBS-T with 1% milk powder in 7 °C with agitation overnight. The antibody solution was decanted and the blot was rinsed briefly twice with TBS-T, then washed three times for 10 min in TBS-T at RT with agitation. The blot was incubated with secondary antibody (Anti-rabbit IgG horse radish peroxidase) diluted to 1:10 000 for 1h at RT in TBS-T with 1% milk-powder with agitation. The blot was washed as above plus once for 10 min with TBS and developed for 5 min with chemiluminescent detection reagent,

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according to manufacture's recommendations. Visualization by exposure to Super RX film (Fujifilm 47410-19236). Exposure time of the film: 30 min.

Courtesy of Dr. Guido Durian, University of Turku, Finland