

This product is for research use only (not for diagnostic or therapeutic use)

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Product no AS09 413

Anti-FtsZ1 and 2 | Plant cell division protein FtsZ1 and FtsZ2

Product information

Immunogen Recombinant part of *Arabidopsis thalina* FtsZ conserved in FtsZ1 Q42545 At5q55280 and FtsZ2 including FtsZ2-1 Q82533, At2q36250 and FtsZ2-2 Q9LXJ0, At3q52750 and in a wide range of FtsZ proteins from other plant species.

Host Rabbit

Clonality Polyclonal

Purity Serum

Format Lyophilized

Quantity 50 ul

Reconstitution For reconstitution add 50 μl of sterile water

Storage Store lyophilized/reconstituted at -20 °C; once reconstituted make aliquots to avoid repeated freeze-thaw cycles. Please remember to spin the tubes briefly prior to opening them to avoid any losses that might occur from material adhering to

the cap or sides of the tube.

Additional information This product can be sold containing proclin if requested

Application information

Recommended dilution 1:2500 (WB)

Expected | apparent

40 kDa FtsZ1 (Arabidopsis thaliana) and 50 kDa FtsZ2 (Arabidopsis thaliana)

Confirmed reactivity Arabidopsis thaliana, Hordeum vulgare, Physcomitrium patens

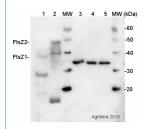
Predicted reactivity Chlamydomons reinhardtii, Cucumis sativus, Gentiana lutea, Glycine max, Gossypium arobretum, Jatropha manihot, Lilium longiflorum, Lupinus angustifolius, Manihot esculenta, Marchantia aquatica, Medicago truncatula, Morus

notabilis, Nannochloropsis gaditana, Nicotiana tabacum, Oryza sativa, Populus trichocarpa, Ricinus communis, Solanum lycopersicum, Sorgum bicolor, Theobroma cacao, Triticum uRatum, Zea mays, Yellow gentian, Vitis vinifera

Species of your interest not listed? Contact us

Not reactive in No confirmed exceptions from predicted reactivity are currently known

Application example



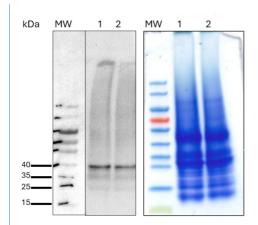
2 μg of total protein extracted with PEB (<u>AS08 300</u>) from leaf tissue of (1) *Hordeum vulgare*, (2) *Arabidopsis thaliana*, (3 - 5) recombinant FtsZ1/FtsZ2 (33 kDa) were separated on 4-12% NuPage (Invitrogen) **LDS-PAGE** and blotted 1h to **PVDF**. Filters were blocked 1h with 2% low-fat **milk powder** in TBS-T (0.1% TWEEN 20) and probed with **anti-FtsZ1/FtsZ2 antibody** (AS09 413, 1:2500, 1h) and secondary anti-rabbit (1:10 000, 1 h) antibody (HRP conjugated, secondary antibody <u>AS09 602</u>) in TBS-T containing 2% low fat milk powder. Antibody incubations were followed by washings in TBS-T (15, +5, +5 min). All steps were performed at RT with agitation. Signal was detected with chemiluminescent detection reagent, using a Fuji LAS-3000 CCD (300s, standard sensitivity). Exposure time was 2 min.



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30 µg of total protein extract from *Physcomitrium patens* protonema culture were loaded as (1) control, and (2) after 2 h treatment with 40 mM glucose. Proteins were separated on a 4–20% TGX gel (Bio-Rad) and transferred to a PVDF membrane for 1 h. Membranes were blocked overnight in TBS-T (0.1% Tween 20) containing 2.5% low-fat milk powder and then probed with anti-FtsZ1/FtsZ2 antibody (AS09 413, 1:2500 in TBS-T with 2.5% low-fat milk) for 1 h at room temperature with agitation. After washing, the membranes were incubated with HRP-conjugated anti-rabbit secondary antibody (AS09 602, 1:25 000 in TBS-T with 2.5% low-fat milk) for 1 h. Washing steps in TBS-T were performed as follows: 3 times for 5 minutes and once for 15 minutes. The signal was developed using chemiluminescent detection reagent (AgriseraECL SuperBright) and visualized with an INTAS ECL ChemoStar. Exposure time was 5 minutes.

Courtesy of Sadia Sayed Tamanna, Technische Universität Kaiserslautern-Landau, Germany