Agrisera
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This product is **for research use only** (not for diagnostic or therapeutic use) **contact: support@agrisera.com**

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Product no AS09 633

Goat anti-Rabbit IgG (H&L), DyLight® 488 conjugated

Product information

Immunogen	Purified Rabbit IgG, whole molecule
Host	Goat
Clonality	Polyclonal
Purity	Immunogen affinity purified goat IgG.
Format	Lyophilized
Quantity	1 mg
Reconstitution	For reconstitution add 1.1 ml of sterile water. Let it stand 30 minutes at room temperature to dissolve. Prepare fresh working dilutions daily
Storage	Store lyophilized material at 2-8°C. Product is stable for 4 weeks at 2-8°C after rehydration. For long time storage after reconstitution, dilute the antibody solution with glycerol to a final concentration of 50% glycerol and store as liquid at -20°C, to prevent loss of enzymatic activity. For example, if you have reconstituted 1 mg of antibody in 1.1 ml of sterile water add 1.1 ml of glycerol. Such solution will not freeze in -20°C, If you are using a 1:5000 dilution prior to diluting with glycerol, then you would need to use a 1:2500 dilution after adding glycerol. Prepare working dilution prior to use and then discard. Be sure to mix well but without foaming.
Additional information	Concentration: 1.0 mg/ml Conjugate is present in 10 mM Sodium Phosphate, 0.15 M Sodium Chloride, pH 7.2, 1 % (w/v) BSA, Protease/IgG free. 0.05 % (w/v) sodium azide is added as preservative. DyLight® 488 has a maximum absorbance at 493 nm; Emax = 518 nm.

Application information

Recommended dilution	1 : 50- 1 : 5 000 (ICC), 1 : 20- 1 : 2000 (IHC), 1 : 3000 (IF)
Confirmed reactivity	Rabbit IgG heavy and light chains (H&L)
Predicted reactivity	Rabbit IgG Heavy and Light chains (H&L)
Additional information	Based in immunoelectrophoresis, this antibody reacts with heavy chains on rabbit IgG and light chains on all rabbit immunoglobulins. No reactivity is observed to non-immunoglobulin rabbit serum proteins based in immunoelectrophoresis. Purity of this antibody is > 95% based on SDS-PAGE.
Selected references	Burchardt et al. (2024). Exploring the response of yellow lupine (Lupinus luteus L.) root to drought mediated by pathways related to phytohormones, lipid, and redox homeostasis. BMC Plant Biol . 2024 Nov 6;24(1):1049. doi: 10.1186/s12870-024-05748-4. Kamińska et al. (2024). Comprehensive elucidation of the differential physiological kale response to cytokinins under in vitro conditions. BMC Plant Biol. 2024 Jul 15;24(1):674.doi: 10.1186/s12870-024-05396-8. Kucko et al. (2022) The acceleration of yellow lupine flower abscission by jasmonates is accompanied by lipid-related events in abscission zone cells, Plant Science, Volume 316, 2022,111173, ISSN 0168-9452, https://doi.org/10.1016/j.plantsci.2021.111173. Namyslov et al. (2020). Exodermis and Endodermis Respond to Nutrient Deficiency in Nutrient-Specific and Localized Manner. Plants (Basel). 2020 Feb 6;9(2). pii: E201. doi: 10.3390/plants9020201. (immunolocalization) Fizesan et al. (2018). Responsiveness assessment of a 3D tetra-culture alveolar model exposed to diesel exhaust particulate matter. Toxicol In Vitro. 2018 Aug 3;53:67-79. doi: 10.1016/j.tiv.2018.07.019. Liu et al. (2016). Fold formation at the compartment boundary of Drosophila wing requires Yki signaling to suppress JNK dependent apoptosis. Sci Rep. 2016 Nov 29;6:38003. doi: 10.1038/srep38003. Wang et al. (2016). Complementary expression of optomotor-blind and the Iroquois complex promotes fold formation to separate wing notum and hinge territories. Dev Biol. 2016 Aug 1;416(1):225-34. doi: 10.1016/j.ydbio.2016.05.020. Epub 2016 May 19

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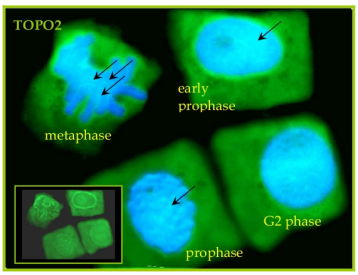
Application example



Seeds of field bean (*Vicia faba* L. subsp. minor var. Nadwiślański; DANKO Group; Sobiejuchy) were sterilized using sodium hypochlorite (0.3% v/v) and germinated in Petri dishes on wetted filter paper at room temperature. At 4 d after imbibition, dark-grown seedlings with primary roots 25±5 mm long were selected for experiments. During incubations roots were oriented horizontally in a humid chamber and aerated continuously on a rotary water-bath shaker (30 rpm) at 23°C. **Immunocytochemical assays** were performed according to the method prescribed earlier (Rybaczek and Maszewski 2006). Excised apical parts of roots (1.5 mm long) were fixed for 45 min (18°C) in PBS-buffered 3.7% paraformaldehyde, washed several times with PBS and placed in a citric acid-buffered digestion solution (pH 5.0; 37°C for 45 min) containing 2.5% pectinase (Fluka), 2.5% cellulase (Onozuka R-10; Serva) and 2.5% pectoliase (ICN). After removing the digestion solution, root tips were washed 3 times in PBS, rinsed with distilled water and squashed onto Super Frost Plus glass slides (Menzel-Gläser). Air-dried slides were pretreated with PBS-buffered 5% BSA at 20°C for 50 min and incubated overnight in a humidified atmosphere (4°C) with rabbit antibody raised against TOPO2 (Agrisera), dissolved in PBS containing 1% BSA (at a dilution of 1:500). Following incubation, slides were washed 3 times with PBS and incubated for 1 h (18°C) with secondary goat anti-rabbit IgG DyLight®488 antibody (Agrisera, AS09 633, 1:3000). Nuclear DNA was stained with 4',6-diamidino-2-phenyl-indole (DAPI, 0.4 µg/ml; Sigma-Aldrich). Following washing with PBS, slides were air dried and embedded in Vectashield Mounting Media for Fluorescence (Vector Laboratories). Observations were made using Optiphot-2 fluorescence microscope (Nikon) equipped with B-2A filter (blue light; 495 nm) for DyLight-conjugated antibodies and UV-2A filter (UV light; 365 nm) for DAPI. All images were recorded at exactly the same time of integration using DXM 1200 CCD camera.

Courtesy Dr. Dorota Rybaczek, Lodz University, Poland