

This product is for research use only (not for diagnostic or therapeutic use)

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### Product no AS21 4605

## Anti-Halo Tag, derived from DhaA of Rhodococcus rhodochrous

### **Product information**

**Immunogen** KLH-conjugated peptide derived from DhaA of *Rhodococcus rhodochrous*, so called HaloTag®.

Host Rabbit

Clonality Polyclonal

**Purity** Antigen affinity purified serum, in PBS pH 7.4

Format Lyophilized

Quantity 50 μg

**Reconstitution** For reconstitution add 50 µl of sterile or deionized water.

Storage Store lyophilized/reconstituted at -20 °C; once reconstituted make aliquots to avoid repeated freeze-thaw cycles.

Please, remember to spin tubes briefly prior to opening them to avoid any losses that might occur from lyophilized material adhering to the cap or sides of the tubes.

Additional information HaloTag® is a trademark of Promega Corporation.

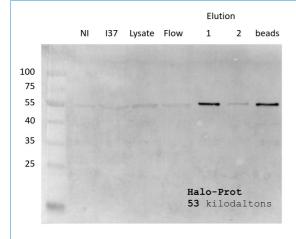
# Application information

Recommended dilution 1:1000 (WB)

Confirmed reactivity Recombinant proteins with HaloTag®

Predicted reactivity Recombinant proteins with HaloTag®

Selected references To be added when available, antibody available in April 2023.



#### Samples:

- 1. Ladder (PageRuler Prestained, 10 à 180 kDa Thermo)
- 2. Non induced bacteria
- 3. Induced bacteria at 37°C for 3h
- 4. Aliquot lysate (after sonication) supernatant
- 5. Flowthrough after incubation with beads
- 6. Elution 1 with acidic elution buffer
- 7. Elution 2 with acidic elution buffer
- 8. Elution of beads with 2x SDS-sample buffer

Recombinant protein was purified from 25 ml E. coli Rossetta bacteria induced at 37 °C during 3 h. After sonication of the culture (lysate), Halo-Trap Agarose beads (chromotek) were added to the supernatant and incubated during 1 h at 4 °C. After 3 wash, beads were eluted with an acidic buffer (x2) and with 50 µl laemmli 2X). Samples were then separated on 12 % SDS-PAGE and blotted for 1 h to nitrocellulose (pore size of 0.45 um), using wet transfer. Blot was blocked with 5% milk for 2 h/RT with agitation. Blot was incubated in the primary antibody at a dilution of 1: 1000 in 5% milk TBS-T ON/4 °C with agitation. The antibody solution was decanted, and the blot was washed 4 times for 5 min in TBS-T at RT with agitation. Blot was incubated with secondary antibody (AS09 602, Rabbit anti-mouse IgG, HRP conjugated) diluted to 1: 20000 in 5% milk for 1h/RT with agitation. The blot was washed as above and developed with a following chemiluminescent detection reagent AS16 ECL-S-10, AgriseraBright. Exposure time was 30 seconds.



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Courtesy of Paul Schouvelier, IBPS, Equipe Biologie de Semences, France