

This product is for research use only (not for diagnostic or therapeutic use)

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Product no AS09 592

Anti-HSP70/HSC70 | Heat shock protein 70

Product information

Immunogen KLH-conjugated C-terminal synthetic peptide conserved in hsc/hsp70 sequences from a wide range of animal species

Host Rabbit

Clonality Polyclonal

Purity Serum

Format Lyophilized

Quantity 200 μl

Reconstitution For reconstitution add 200 μl of sterile water

Storage Storage Store lyophilized/reconstituted at -20 °C; once reconstituted make aliquots to avoid repeated freeze-thaw cycles. Please remember to spin the tubes briefly prior to opening them to avoid any losses that might occur from material adhering to

the cap or sides of the tube.

Additional information Chosen peptide sequence is also conserved in several fish species including: <u>DQ202278.1</u> HSC70 Fundulus,

DQ202279.1 HSP70-1 Fundulus, DQ202280.1 HSP70-2 Fundulus, BT059361.1 HSC70 Salmo salar (atlantic salmon), AB092839.2 HSP70 Carassius auratus (goldfish), BC056709.1 HSP70 Danio rerio (zebrafish) and other animal HSP70

proteins

Application information

Recommended dilution 1:10 000 (WB)

Expected | apparent 70 kDa

MW 70 KL

Confirmed reactivity Arabidopsis thaliana, Acanthamoeba castellanii (amoeba), Caenorhabditis elegans, salmon (Salmo salar),

Dictyostelium discoideum, frog-heart, Frog-skeletal muscle, Frog-liver, rainbow trout (Oncorhynchus mykiss), cow,

Chicken, pig, Rat, seal, mummichog

Predicted reactivity Gammarus pulex, Salmo salar (Atlantic salmon), Salvelinus fontinalis (Brook trout)

Species of your interest not listed? Contact us

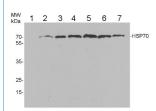
Not reactive in No confirmed exceptions from predicted reactivity are currently known

Selected references Peláez et al. (2025). Harnessing macroalgal cell walls to trigger immunity in Arabidopsis thaliana. Plant Stress Volume

15, March 2025, 100783.

Chandra et al. (2012). Sustained high temperature increases the vitellogenin response to 17 alpha-ethynylestradiol in mummichog (Fundulus heteroclitus). Aquatic toxicology.

Application example



Dictyostelium discoideum (1), Acanthamoeba castellanii (2), Frog-heart (3), Frog-skeletal muscle (4), Frog-liver (5), Caenorhabditis elegans (6), Arabidopsis thaliana (7), tissues were homogenized in glass homogenizer in PBS buffer and centrifuged at 500 x g for 5 min. Supernatant was collected and 50 μg of protein for each gel lane was denatured with Laemmli buffer at 950C for 5 min. Samples were separated on 14 % SDS-PAGE and blotted 1h to nitrocellulose using semi-dry transfer. Blot was blocked overnight in 5% milk in TBS buffer and next incubated in the primary antibody at a dilution of 1: 2000 for 1h at RT with agitation. The antibody solution was decanted and the blot was washed once for 15 min and 4 times for 5 min in TBS-T at RT with agitation. Blot was incubated in secondary antibody (anti-rabbit IgG horse radish peroxidase conjugated, from) diluted to 1:25000 for 1h at RT with agitation. The blot was washed as above and developed for 5 min with chemiluminescent detection. Exposure time was 300 seconds.

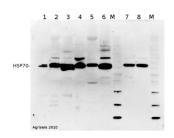
Courtesy Dr. Małgorzata Słocińska, UAM, Poland



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5 μg of total protein from (1) cow muscle, (2) chicken muscle, (3) pig muscle, (4) rat liver, (5) salmon muscle, (6) seal muscle, (7) mummichog heat shock control, (8) mummichog heat shock post-24 hours, extracted with Protein Extration Buffer, PEB (AS08 300), were separated on 4-12% NuPage (Invitrogen) LDS-PAGE and blotted 1h to PVDF. Blots were blocked immediately following transfer in 2% ECL Advance blocking reagent (GE Healthcare) in 20 mM Tris, 137 mM sodium chloride pH 7.6 with 0.1% (v/v) Tween-20 (TBS-T) for 1h at room temperature with agitation. Blots were incubated in the primary antibody at a dilution of 1: 10 000 for 1h at room temperature with agitation. The antibody solution was decanted and the blot was rinsed briefly twice, then washed once for 15 min and 3 times for 5 min in TBS-T at room temperature with agitation. Blots were incubated in secondary antibody (Agrisera anti-rabbit IgG horse radish peroxidase conjugated, AS09 602) diluted to 1:50 000 in 2% ECL Advance blocking solution for 1h at room temperature with agitation. The blots were washed as above and developed for 5 min with ECL Advance detection reagent according to the manufacturers instructions. Images of the blots were obtained using a CCD imager (FluorSMax, Bio-Rad) and Quantity One software (Bio-Rad). Exposure time was 30 seconds.