Agrisera
Part of Olink® Group

This product is **for research use only** (not for diagnostic or therapeutic use) **contact: support@agrisera.com**

Agrisera AB | Box 57 | SE-91121 Vännäs | Sweden | +46 (0)935 33 000 | www.agrisera.com

Product no AS16 3688

JAR1 | Jasmonic acid-amido synthetase JAR1

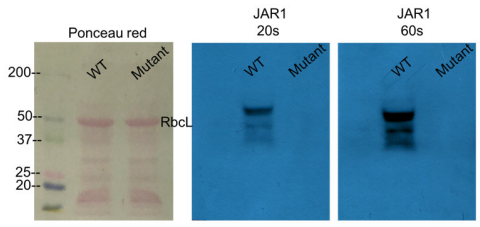
Product information

Immunogen	KLH-conjugated synthetic peptide derived from *Arabidopsis thaliana* JAR1 protein sequence, UniProt: Q9SKE2, TAIR: At2g46370
Host	Rabbit
Clonality	Polyclonal
Purity	Immunogen affinity purified serum in PBS pH 7.4.
Format	Lyophilized
Quantity	50 µg
Reconstitution	for reconstitution add 50 µl, of sterile water
Storage	Store lyophilized/reconstituted at -20°C; once reconstituted make aliquots to avoid repeated freeze-thaw cycles. Please remember to spin the tubes briefly prior to opening them to avoid any losses that might occur from material adhering to the cap or sides of the tube.

Application information

Recommended dilution	1: 1000 (WB)
Expected | apparent MW	64 kDa (*Arabidopsis thaliana*)
Confirmed reactivity	*Arabidopsis thaliana*
Predicted reactivity	*Capsicum annuum, Gossypium hirsutum, Nicotiana tabacum, Prunus yedoensis var. nudiflora, Solanum tuberosum, Ulmus americana* Species of your interest not listed? Contact us
Not reactive in	
Additional information	Phenol protein extraction method protocol.

Application example



5 µg of protein from *Arabidopsis thaliana* Col0 and *jar1-1* mutant were extracted using basic phenol protocol. 1g were grinded with liquid nitrogen with mortair and pestle, powder were resuspended in 3 mL of protein buffer (0.5 M Tris-HCl, 0.7 M sucrose, 1 mM PMSF, 50 mM EDTA, 0.1 M KCl and 0.2% -mercaptoethanol; pH 8.0). Homogenate were centifuged at 8000 rpm and supernatant were mixed with three volumes of basic phenol, the mixture were incubated with agitation at room temperature during 10 mins. The mixture were centrifuged at 10000 rpm at 4ºC during 30 min, phenolic phase were recuperated in new tube and were mixed with one volume of ammonium acetate 0.1M in methanol, the mixture were incubated at -20 ºC during 4 hours. The mixture were centrifuged at 10000 rpm at 4º C during 20 minutes and protein pellet were washed three times with ammonium acetate 0.1M in methanol. Finally proteins were resuspended in tris-HCl (50 mM, pH 8.0) and the protein concentration were determined according Bradford method. Samples of 5 and 10µg were separated on 15 % SDS-PAGE using tank (BioRad system) to transfer to nitrocellulose membrane during 1 hour at 400 A. Blots were blocked with skimmed milk (10%) for 1h at room temperature (RT) with agitation. Blot was incubated in the primary antibody at a dilution of 1: 1000 for 1h at RT with agitation. The antibody solution was decanted and the blot was rinsed briefly twice, then washed once for 15 min and 3 times for 5 min in TBS-T at RT with agitation. Blot was incubated in secondary antibody (anti-rabbit IgG horse radish peroxidase conjugated, from Agrisera AS09 602) diluted to 1:20 000 in for 1h at RT with agitation. The blot was washed as above and developed for 5 min with ECL according to the manufacturer's instructions. Exposure time was 30 and 60 seconds. For transference control the membrane was stained with Ponceau red and integrity of proteins was evaluated using 12% SDS-PAGE comassie blue stained.

Courtesy of Dr. Rodrigo Contreras, Universidad de Santiago de Chile, Chile