

This product is for research use only (not for diagnostic or therapeutic use)

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Product no AS16 3221-1ml

Anti-Non-fucosylated xyloglucan-5 (clone CCRC-M48)

Product information

 Immunogen
 BSA-conjugated tamarind xyloglucan (covalent). Epitope structure for carbohydrate antigen: XXLG, XLLG.

 Host
 Mouse

 Clonality
 Monoclonal

 Subclass/isotype
 IgG1

 Purity
 Cell culture supernatant.

 Format
 Liquid

 Quantity
 1 ml

 Storage
 Antibody can be stored up to 1 month at 4°C, and at -80°C for up to 1 year. Make aliquots to avoid repeated freeze-thaw cycles. Please remember to spin the tubes briefly prior to opening them to avoid any losses that might occur from material adhering to the cap or sides of the tube.

 Additional information
 Exact working dilution needs to be determined by end user

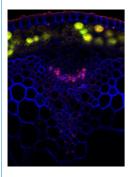
 Application information
 Undiluted or 1 : 10 (ELISA), (IHC), (IF)

 Confirmed reactivity
 Arabidopsis thaliana, Camelina sativa, Tamarindus indicus

 Predicted reactivity
 Dicots

Predicted reactivity	Dicots Species of your interest not listed? <u>Contact us</u>
Not reactive in	No confirmed exceptions from predicted reactivity are currently known
Additional information	CCRC-M48 binds to galactosylated side-chains of non-fucosylated xyloglucan, and appears to preferentially bind to the galactosylated side-chain closest to the reducing end of xyloglucan oligosaccharide sub-units (XXLG, XLLG)

Application example



Localization of non-fucosylated xyloglucan-5 (red) in Arabidopsis thaliana hypocotyl, Calcufluor White counterstain (blue) and cell wall autofluorescence (yellow).

The 31 days-old hypocotyls were immersed in 150 μ L PME fixation buffer (25 mM PIPES, 1 mM MgSO₄, 1 mM EGTA) and then subjected to three consecutive cycles of 5 min-long vacuum infiltration (21°C, 68 kPa). Afterwards they were washed three times in PME (21°C, 68 kPa) prior to storage at 4°C in PME. Hypocotyls were encased in 1 cm³ blocks of 5% agar at 65°C, and stored at 4°C to set. Transverse 40 μ m thick sections were cut from segments using a VT100S vibrating microtome (Leica) and blocked for at least 1 h in 5% bovine serum albumin in TBST. Blocking solution was discarded and sections were incubated at 4°C for 16 h with 5 μ l of the anti-Non-fucosylated xyloglucan-5 antibody, followed by 2 washes in 100 μ L TBST. Sections were then incubated for 1 h at 21°C in the dark in 10 μ l of 2 μ g/ μ l Alexa FluorTM 568 donkey anti-mouse IgG (H+L; 1:36). Sections were again washed twice in 40 μ L TBST prior to counter-staining with 0.015% Calcofluor White (Sigma-Aldrich). Sections were again washed twice in 100 μ L TBST to remove excess counter-stain and unbound secondary antibody. Immunofluorescence of AlexaFluor 568 was excited with a 561 nm laser, and emitted light filtered at 575–600 nm, while Calcufluor White was subsequently scanned on an independent channel with a 405 nm laser and emission observed at 420–430 nm using laser scanning microscope Zeiss LSM780 point-scan system at 1024 × 1024 pixels (pixel size, 0.6–0.83 μ m) with a 10X objective.

Courtesy Dr. Urs Fisher, Umeå Plant Science Centre, Sweden