

This product is **for research use only** (not for diagnostic or therapeutic use)

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Product no AS16 3225-1ml

Anti-Rhamnogalacturonan-I / Arabinogalactan (clone CCRC-M7)

Product information

Immunogen MeBSA-conjugated sycamore rhamnogalacturonan-I (non-covalent),

Host Mouse

Clonality Monoclonal

Subclass/isotype | IgG1

Purity Cell culture supernatant.

Format Liquid

Quantity 1 ml

Storage

Antibody can be stored up to 1 month at 4°C, and at -80°C for up to 1 year. Make aliquots to avoid repeated freeze-thaw cycles. Please remember to spin the tubes briefly prior to opening them to avoid any losses that might occur from material adhering to the cap or sides of the tube.

Additional information

Exact working dilution needs to be determined by end user, Epitope structure for carbohydrate antigen: trimer or larger of beta-(1,6)-Gal carrying one or more Ara residues of unknown linkage

Application information

Recommended dilution Undiluted or at 1:10 (ELISA), (IHC), (IF)

Predicted reactivity Dicots

Species of your interest not listed? Contact us

Not reactive in No confirmed exceptions from predicted reactivity are currently known

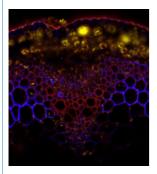
Selected references Pattathil et al. (2012). Immunological approaches to plant cell wall an

Pattathil et al. (2012). Immunological approaches to plant cell wall and biomass characterization: Glycome Profiling.

Methods Mol Biol. 2012;908:61-72.doi: 0.1007/978-1-61779-956-3_6.

Patathil et al. (2010). A comprehensive toolkit of plant cell wall glycan-directed monoclonal antibodies. Plant Physiol. 2010 Jun;153(2):514-25.doi: 10.1104/pp.109.151985.

Application example



Localization of Rhamnogalacturonan-I/Arabinogalactan (red) in *Arabidopsis thaliana* hypocotyl, Calcufluor White counterstain (blue) and cell wall autofluorescence (yellow).

The 31 days-old hypocotyls were immersed in 150 μ L PME fixation buffer (25 mM PIPES, 1 mM MgSO₄, 1 mM EGTA) and then subjected to three consecutive cycles of 5 min-long vacuum infiltration (21 °C, 68 kPa). Afterwards they were washed three times in PME (21 °C, 68 kPa) prior to storage at 4 °C in PME. Hypocotyls were encased in 1 cm³ blocks of 5% agar at 65 °C, and stored at 4 °C to set. Transverse 40 μ m thick sections were cut from segments using a VT100S vibrating microtome (Leica) and blocked for at least 1 h in 5% bovine serum albumin in TBST. Blocking solution was discarded and sections were incubated at 4 °C for 16 h with 5 μ l of the anti-Rhamnogalacturonan-l/Arabinogalactan antibody, followed by 2 washes in 100 μ L TBST. Sections were then incubated for 1 h at 21 °C in the dark in 10 μ l of 2 μ g/ μ l Alexa FluorTM 568 donkey anti-mouse IgG (H+L; 1:36). Sections were again washed twice in 40 μ L TBST prior to counter-staining with 0.015% Calcofluor White (Sigma-Aldrich). Sections were again washed twice in 100 μ L TBST to remove excess counter-stain and unbound secondary antibody. Immunofluorescence of AlexaFluor 568 was excited with a 561 nm laser, and emitted light filtered at 575–600 nm, while Calcufluor White was subsequently scanned on an independent channel with a 405 nm laser and emission observed at 420–430 nm using laser scanning microscope Zeiss LSM780 point-scan system at 1024 × 1024 pixels (pixel size, 0.6–0.83 μ m) with a 10X objective.

Courtesy Dr. Urs Fisher, Umeå Plant Science Centre, Sweden