

This product is for research use only (not for diagnostic or therapeutic use)

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Product no AS21 4508 Anti-Trx Tag | Thioredoxin 1 Fusion protein

Product information

Immunogen	Full-lenght recombinat Trx protein, UniProt: POAA25 expressed in E.coli
Host	Rabbit
Clonality	Polyclonal
Purity	Antigen affinity purified serum, in PBS pH 7.4
Format	Lyophilized
Quantity	50 μg
Reconstitution	For reconstitution, add 50 μ l of sterile or deionized water.
Storage	Store lyophilized/reconstituted at -20°C; once reconstituted make aliquots to avoid repeated freeze-thaw cycles. Please, remember to spin tubes briefly prior to opening them to avoid any losses that might occur from lyophilized material adhering to the cap or sides of the tubes.

Application information

Recommended dilution	1 : 2000 - 1: 5000 (WB)
Expected apparent MW	11.8 kDa (Trx tag)
Confirmed reactivity	Trx Fusion protein
Not reactive in	No confirmed exceptions from predicted reactivity are currently known
Selected references	To be added when available, antibody available in April 2025.



Analysis of production of protein of interest (POI): H10-TRX-POIoverexpressed in E. Coli BL21 (DE3) Rosetta

Samples:

- 1- 10 μl of total fraction of cell lysate, before induction.
- 2- 10 µl of total fraction of cell lysate of IPTG (0.05mM) induced (ON, 16ºC) BL21 (DE3) Rosetta.
- 3- 10 µl of soluble fraction of cell lysate of IPTG (0.05mM) induced (ON, 16ºC) BL21 (DE3) Rosetta.
- 4- 10 μl of insoluble pellet fraction of cell lysate of IPTG (0.05mM) induced (ON, 16°C) BL21 (DE3) Rosetta.
- 5- 10 µl of total fraction of cell lysate of IPTG (0.1mM) induced (ON, 16ºC) BL21 (DE3) Rosetta.
- 6- 10 µl of soluble fraction of cell lysate of IPTG (0.1mM) induced (ON, 16ºC) BL21 (DE3) Rosetta.
- 7- 10 µl of insoluble pellet fraction of cell lysate of IPTG (0.1mM) induced (ON, 16ºC) BL21 (DE3) Rosetta.
- 8- 10 µl of soluble fraction of negative control H10-POI

10 μl of protein extract from BL21 (DE3) Rosetta (all bacteria pellets were normalized to final DO of 6.5). Exact buffer components were: Tris-HCl 50 mM, NaCl 500mM, 10% glycerol, pH:8 and denatured with 2X buffer (Tris-HCl 125 mM, 20% glycerol, SDS 4%, BeOH, 2% and blue bromophenol 0.001%) at 95 °C 5min. Samples were separated in the cold on 10 % SDS-PAGE and blotted for 1h 30min nitrocellulose (0.45 μm), using: wet transfer in the cold. Blot was blocked with 5 % milk for: 2h/RT with agitation. Blot was incubated in the primary antibody at a dilution of 1: 2000 for ON/4 °C with agitation. The antibody solution was decanted, and the blot was rinsed briefly twice, then washed 3 times for 10 min in TBS-T at RT with agitation. Blot was incubated in matching secondary antibody (anti-rabbit IgG horse radish peroxidase conjugated) diluted to 1:10 000 in 5% nonfat milk TBT-T for 1 h/RT with agitation. The blot was washed as above and developed with a following chemiluminescent detection reagent. Exposure time was 30 seconds.

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