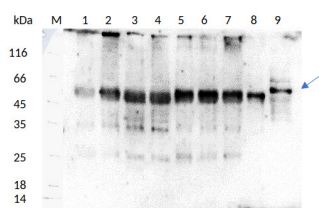


Product no **AS20 4482****Anti-Tubulin gamma chain (monoclonal antibodies)****Product information**

<b>Immunogen</b>	KLH-conjugated gamma-tubulin peptide EYHAATRPDYISWGTQ, amino acids 434-449. UniProt: <a href="#">P23258</a> The epitope was located in the amino acid sequence PDYISW (aa441-446 in human), which is identical for gamma-tubulin 1 and gamma-tubulin 2.
<b>Host</b>	Mouse
<b>Clonality</b>	Monoclonal
<b>Subclass/isotype</b>	IgG1
<b>Purity</b>	Immunoglobulin Protein A purified in PBS. Contains 15 mM sodium azide.
<b>Format</b>	Liquid
<b>Quantity</b>	100 µg
<b>Storage</b>	Store at 4°C; Do not freeze. Do not exceed expiry date is provided on the tube. Please remember to spin the tubes briefly prior to opening them to avoid any losses that might occur from material adhering to the cap or sides of the tube.

**Application information**

<b>Recommended dilution</b>	2-8 µg/ml (ICC) methanol/acetone fixation required, 1-2 µg/ml (WB)
<b>Expected   apparent MW</b>	51 kDa
<b>Confirmed reactivity</b>	<i>Chlorella vulgaris</i> , <i>Nicotiana tabacum</i> , human, protozoa
<b>Predicted reactivity</b>	Species of your interest not listed? <a href="#">Contact us</a>
<b>Not reactive in</b>	No confirmed exceptions from predicted reactivity are currently known
<b>Additional information</b>	This antibody recognizes C-terminus (amino acids 434-449 in human) of gamma-tubulin, a 48 kDa structural constituent of cytoskeleton and microtubule organizing center (MTOC). The epitope which this antibody is recognizing is conserved in <i>Arabidopsis thaliana</i> Tubulin gamma-1 chain, UniProt: <a href="#">P38557</a> , Gene ID: <a href="#">At3g61650</a> and Tubulin gamma-2 chain, UniProt: <a href="#">P38558</a> , Gene ID: <a href="#">At5g05620</a>  Recommended secondary antibody: goat anti-mouse IgG1 <a href="#">AS16 3715</a>
<b>Selected references</b>	<a href="#">Romeiro Motta et al. (2024)</a> . The cell cycle controls spindle architecture in Arabidopsis by activating the augmin pathway. Dev Cell. 2024 Aug 23:S1534-5807(24)00484-2. doi: 10.1016/j.devcel.2024.08.001.

**Samples:**

- 1 - 14 µg of *Nicotiana tabacum*, L. whole leaf extract of adult plant- control (untreated)
- 2 - 14 µg of *Nicotiana tabacum*, L. whole leaf extract of adult plant after treatment with 100 µM AgNP-PVP
- 3 - 14 µg of *Nicotiana tabacum*, L. whole leaf extract of adult plant after treatment with 100 µM AgNP-CTAB
- 4 - 14 µg of *Nicotiana tabacum*, L. whole leaf extract of adult plant after treatment with 100 µM AgNO<sub>3</sub>
- 5 - 14 µg of *Nicotiana tabacum*, L. whole leaf extract of adult plant after treatment with 100 µM AgNP-PVP+cys
- 6 - 14 µg of *Nicotiana tabacum*, L. whole leaf extract of adult plant after treatment with 100 µM AgNP-CTAB+cys
- 7 - 14 µg of *Nicotiana tabacum*, L. whole leaf extract of adult plant after treatment with 100 µM AgNO<sub>3</sub>+cys
- 8 - 14 µg of *Nicotiana tabacum*, L. extract of 4 weeks old seedlings- control (untreated)
- 9 - 14 µg of *Chlorella vulgaris* extract- control (untreated)

14 µg/well of total protein extracted freshly from 0.05 g of lyophilized whole leaf and root of adult tobacco plants following the Phenol extraction protocol1 with extraction buffer containing Trizma base (500 mM), Ethylenediaminetetraacetic acid (EDTA) (50 mM), sucrose (700 mM) and Potassium chloride (KCl) (100 mM) with addition of phenylmethylsulfonyl fluoride (PMSF) (1 mM) and 2% -mercaptoethanol. After short incubation on 4°C with agitation, phenol was added. The phenol (supernatant) phase containing proteins, was collected after centrifugation and equal volume of extraction buffer was added. After centrifugation, supernatant phase was collected and 4 volumes of 0.1 M ammonium acetate

(with 10% methanol) was added, and proteins were precipitated ON/-20 °C. The next day, protein pellets were washed 3 times in ammonium acetate with rounds of centrifugations in between, and finally in acetone. Protein pellet was lastly resuspended in Isoelectric focusing buffer (IEF) containing 9 M urea, 4% CHAPS, 20 mM DTT, 1.2% Ampholytes pH 3 to 10. Protein concentrations were measured with modified Bradford method<sup>1</sup> and denatured with Laemmli sample buffer<sup>2</sup> at 95 °C for 5 min. Total proteins were separated on 12 % SDS-PAGE and blotted 1h to nitrocellulose (pore size of 0.2 µm), using wet transfer. Blot was blocked with 2 % milk in PBS-T, 1h/RT. Blot was incubated in the primary antibody at a dilution of 1: 1 000 for 1h/RT with agitation in a solution of 2 % milk in PBS-T and then ON/4 °C. The antibody solution was decanted and the blot was then washed 3 times for 10 min in 2% milk in PBS-T at RT with agitation. Blot was incubated in Agrisera matching secondary antibody (goat anti-Mouse IgG1 HRP conjugated, [AS16 3715](#)) diluted to 1:10 000 in 2% milk in PBS-T for 1h/RT with agitation. The blot was washed twice for 10 min in PBS-T developed for 5 min with AgriseraECLSuperBright ([AS16 ECL-S](#)). Exposure time was 12 min.

Courtesy of MSc, Karla Košpić, University of Zagreb, Faculty of Science Department of Biology, Croatia