

This product is for research use only (not for diagnostic or therapeutic use)

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## Product no AS22 4716

## Anti-UVR8 | Ultraviolet-B receptor UVR8

## **Product information**

Immunogen KLH-conjugated peptide derived from Arabidopsis thaliana UVR8, UniProt: Q9FN03, TAIR: At5g63860

**Host** Rabbit

Clonality Polyclonal

**Purity** Antigen affinity purified serum, in PBS pH 7.4

Format Lyophilized

Quantity 50 μg

**Reconstitution** For reconstitution, add 50 μl, of sterile or deionized water.

Store lyophilized/reconstituted at -20°C; once reconstituted make aliquots to avoid repeated freeze-thaw cycles. Please, remember to spin tubes briefly prior to opening them to avoid any losses that might occur from lyophilized

material adhering to the cap or sides of the tubes.

## Application information

Recommended dilution 1:1000 (WB)

Expected | apparent

47.12 kDa

Predicted reactivity

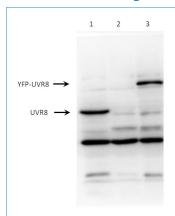
Brassica napus. Coffea arabica, Capsicum annuum, Glycine soja, Gossypium australe, Hordeum vulgare, Ipomoea

triloba, Malus domestica, Nicotiana benthamiana, Nicotiana tabacum, Solanum lycopersicum, Solanum tuberosum, Oryza sativa, Phtheirospermum japonicum, Populus alba x Populus x berolinensis, Senna tora, Triticum aestivum,

Triticum urartu, Turnera subulata, Zea mays Species of your interest not listed? Contact us

**Not reactive in** No confirmed exceptions from predicted reactivity are currently known

**Selected references** To be added when available, antibody available in May 2023.



Each lane contains 20 µg of *Arabidopsis thalian*a whole leaf extract of the following genetic backgrounds:

- 1. Columbia ecotype (UVR8 wild type)
- 2. uvr8-6 mutant
- 3. uvr8-6/proUVR8:YFP-UVR8 (described in Bernula et al. 2017)

20 µg/well of total protein extracted from liquid N2-frozen leaves of adult plants using hot extraction buffer containing 65 mM Tris-HCl, pH 7.8, 4 M urea, 5% (w/v) SDS, 100 mM dithiothreitol, 15% (v/v) glycerol, and 0.05% (w/v) bromophenol blue and denatured at 95°C for 5 min. Samples were separated at RT on 10% SDS-PAGE and blotted for 2 h to PVDF (pore size of 45 µm), using wet transfer in the cold. Blot was blocked with 5% milk 1h/RT with agitation. The blot was incubated in the primary antibody at a dilution of 1:1000 in TBS-T during overnight (16 h) at 4 °C with agitation. The antibody solution was decanted, and the blot was rinsed briefly, then washed 5 times for 5 min in TBS-T at RT with agitation. Blot was incubated in matching secondary antibody (anti-rabbit IgG horse radish peroxidase conjugated, AS09 602 Agrisera) diluted to 1:10 000 in TBS-T for 1 h/RT with agitation. The blot was washed as above and developed with a following chemiluminescent detection reagent: AS16 ECL-S-10 AgriseraSuperBright. Exposure time was 0.6 s (iBright, 3x3 binning).

Courtesy of Dr. András Viczián, Institute of Plant Biology. Biological Research Center Szeged Hungary