

This product is for research use only (not for diagnostic or therapeutic use)

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## Product no AS14 2775

## Anti-V-ATPase, B | vacuolar H+-ATPase subunit B

## **Product information**

Immunogen KLH-conjugated synthetic peptide chosen from Arabidopsis thaliana V-ATPase subunit B, isoform

B1: UniProt: Q683E8, TAIR: AT1G76030, isoform B2 UniProt: Q9SZN1, TAIR: AT4G38510, isoform B3: UniProt:

Q8W4E2, TAIR: AT1G20260

**Host** Rabbit

Clonality Polyclonal

**Purity** Immunogen affinity purified serum in PBS pH 7.4.

Format Lyophilized

Quantity 50 μg

**Reconstitution** For reconstitution add 50 µl of sterile water

Store lyophilized/reconstituted at -20 °C; make aliquots to avoid repeated freeze-thaw cycles. Please remember to spin

the tubes briefly prior to opening them to avoid any losses that might occur from material adhering to the cap or sides of

the tube.

**Additional information** Antibodies will detect target protein in a few µg of a crude preparation loaded per well, If purified preparations of

vacuolar and plasma membranes are used, one μg load per well should be sufficient

# Application information

Recommended dilution 1:1000 (WB)

Expected | apparent

53 | 57 kDa (Vigna radiata)

Confirmed reactivity Arabidopsis halleri, Nicotiana tabaccum, Thellungiella salsuginea, Vigna radiata

Predicted reactivity

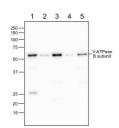
Acetabularia sp., Arundo donax, Chlamydomonas reinhardtii, Cucumis sativus, Glycine soja, Gossypium mexicanum, Halostachys caspica, Haloxylon ammodendron, Hordeum vulgare, Medicago truncatula, Mesembryantheum crystallinum, Ostreococcus tauri, Oryza sativa, Panax ginseng, Physcomitrium patens, Pinus sylvestris, Populus trichocarpa, Pyrus sp., Ricinus communis, Theobroma cacao, Triticum aestivum, Zea mays, Zostera marina

Species of your interest not listed? Contact us

Not reactive in Thermatoga neapolitana

Additional information Protein or membrane sample should be treated at 70°C for 10 min before loading on the gel

### Application example



Proteins were separated by SDS-PAGE and transferred to an Immobilon-P membrane (Millipore) using Trans-Blot SD Semi-Dry Transfer Cell (Bio-Rad) with transfer buffer (100 mM Tris, 192 mM Glycine, 0.02% (w/v) SDS and 5% (v/v) methanol). After treatment with 1% blocking agent, the membrane filter was incubated with the primary antibody (1:1000) and then with horseradish peroxidase (HRP)-conjugated goat anti-rabbit IgG (H+L) (Agrisera AS09 602, 1:25 000). Chemiluminescent reagent was used for detection of antigens. Chemiluminescence was detected with a Light-Capture II imaging device with a cooled CCD camera (Atto).

- 1: 10 μg of 100,000 x g precipitate prepared from *Arabidopsis thaliana* 6 weeks old shoot.
- 2: 0.2 µg of vacuolar membrane enriched fraction prepared from Arabidopsis thaliana 6 weeks old shoot.
- 3: 2 µg of vacuolar membrane enriched fraction prepared from *Arabidopsis thaliana* 6 weeks old shoot.
- 4: 0.2 μg of vacuolar membrane enriched fraction prepared from Vigna radiata 4 days old hypocotyls. 5: 2 μg of vacuolar membrane enriched



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fraction prepared from Vigna radiata 4 days old hypocotyls.

Courtesy of Drs. Masayoshi Maeshima and Dr Shoji Segami, Nagoya University, Japan