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Product no AS07 267

Anti-Xylose

Product information

Immunogen	xylose residues bound to the N-glycan in beta 1,2
Host	Rabbit
Clonality	Polyclonal
Purity	Immunogen affinity purified serum in PBS pH 7.4.
Format	Lyophilized
Quantity	50 μg
Reconstitution	For reconstitution add 50 µl of sterile water
Storage	Store lyophilized/reconstituted at -20°C; once reconstituted make aliquots to avoid repeated freeze-thaw cycles. Please remember to spin the tubes briefly prior to opening them to avoid any losses that might occur from material adhering to the cap or sides of the tube.
Additional information	Beta (1,2) xylose is present exclusively in plant N-glycans so antibodies against this sugar moiety should not cross-react with any mammal glycoprotein.
	This antibody do not bind free D-xylose. This antibody does not seem to work in immunolocalization.

Application information

Recommended dilution	1 μg/ml (ELISA), 2 μg/10 ml incubation buffer (WB)
Expected apparent MW	10-100 for various glycoproteins
Confirmed reactivity	Higher plants and algae
Predicted reactivity	Higher plants
Not reactive in	No confirmed exceptions from predicted reactivity are currently known
Additional information	Negative control: Fetuin, a glycoprotein containing fucose linked in alpha 1.6 and no xylose, Sigma, product number F3385. Positive control: Type II - horseradish peroxidase which contains 1.2 Xylose and 1.3 fucose, Sigma, product number
	P8250
Selected references	Xu et al. (2023). The Golgi-localized transporter OsPML4 contributes to manganese homeostasis in rice. Plant Sci. 2023 Dec 2:339:111935.doi: 10.1016/j.plantsci.2023.111935. Ropitaux et al. (2023). Subcellular localization of core beta (1, 2)-xylosylated N-glycoproteins in the green microalgae Chlamydomonas reinhardtii. Algal Research Volume 77, January 2024, 103366. Xavier et al. (2021) Inactivation of N-Acetylglucosaminyltransferase I and a1,3-Fucosyltransferase Genes in Nicotiana tabacum BY-2 Cells Results in Glycoproteins With Highly Homogeneous, High-Mannose N-Glycans. Frontiers in Plant Science. Volume 12. doi: 10.3389/fpls.2021.634023 Yang et al. (2021). Golgi-localized manganese transporter PML3 regulates Arabidopsis growth through modulating Golgi glycosylation and cell wall biosynthesis. New Phytol. 2021 Jan 17. doi: 10.1111/nph.17209. Epub ahead of print. PMID: 33454966. Lucas et al. (2019). Multiple xylosyltransferases heterogeneously xylosylate protein N-linked glycans in Chlamydomonas reinhardtii. Plant J. 2019 Nov 27. doi: 10.1111/tpj.14620. Lucas et al. (2019). Multiple xylosyltransferases heterogeneously xylosylate protein N-linked glycans in Chlamydomonas reinhardtii. Plant J. 2019 Nov 27. doi: 10.1111/tpj.14620. Lucas et al. (2019). Identification and characterization of a novel glycoprotein core xylosidase from the bacterium Elizabethkingia meningoseptica. Biochemical and Biophysical Research Communications Available online 27 July 2019.

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mc	eau r	eu s	tain	ing				An	Anti-Xylose antibody AS07 267 lot n°								
	MW	1	2	3	4	5	6		MW	1	2	3	4	5	6		
250								250									
30	-				1			130	-								
00	-				1			100	-	51							
0	-				1			70	-	H							
5	-				1			55	-	_							
		П			I					1							
15	-	E	Ξ		1			35	-			5					
5	-							25		1		1					
5	-							15 10	-								

- 1 50 ug of cytosolic proteins from Arabidopsis thaliana WT
- 2 50 ug of cytosolic proteins from Arabidopsis thaliana fut11/fut12 mutant
- 3 50 ug of cytosolic proteins from Arabidopsis thaliana xylt mutant
- 4-50 ug of cytosolic proteins from Arabidopsis thaliana xylt/ fut11/fut12 mutant
- 5-1 ug of recombinant Avidin produced in maize (SIGMA)
- 6-1 ug of Bovine Ribonuclease B (SIGMA) Mark: MW markers: PAGE Ruler pre-stained protein ladder from Fisher scientific

Protein extractions of about 500 mg of leaf of *Arabidopsis thaliana* in 1 mL Tris Buffer 0.1 M pH 7.5 were carried out in a high-speed benchtop homogenizer (FastPrep®-24 MP) 6 m.s-1, 40 s, 2 times, followed by a centrifugation step for 5 min at 10 000 rpm, 4°C. The pellet is discarded while the supernatant contains what we called cytosolic proteins. 20 μ g of cytosolic proteins (WT and mutants Fut11/Fut12 ; xyltT ; xylt/fut11/Fut12) denaturated with learnmli buffer at 100°C for 5 min were separated on Novex Bis-Tris 4-12% gel in MOPS buffer, and blotted on nitrocellulose using semi-dry transfer Thermo ScientificTM PierceTM Power Blotter for 13 min in one step transfert buffer (thermoScientific 84731). Incubation of the blot is carried out with SNAP i.d.@ 2.0 Protein Detection System according to the technical specifications of the supplier. Quickly, blot was blocked in TBST for 10 min, then the vacuum was applied. The primary antibody Anti- (1,2)-Xylose (AS07 267) was used at a dilution of 1:3000 in TBST and left on the blot for 15 min before vacuum was applied. Three washes with TBST were performed and the secondary antibody (Goat anti rabbit IgG-HRP conjugated <u>AS09 602</u>) was used at a dilution 1:50 000 in TBST and left for 15 min with Agrisera ECL set (<u>AS16 ECL-SN-10</u>) and time exposure is 30 sec performed with the VILBER Fusion FX Imager. We used Avidin (1 μ g) as positive control and Ribonuclease B (1 μ g) as negative control. Avidin carries 1,2 Xylose and ribonuclease B only mannose.

Conclusion: The Anti-(1,2)-Xylose antibody (AS07 267) lot n° 2108 as expected, shows a positive response with Avidin, WT and fuT mutant and a negative response with the XylT mutant of Arabidopsis thaliana. It exhibits a negative response with ribonuclease B as expected too. Thus demonstrate the good specificity of the Anti-(1,2)-Xylose antibody (AS07 267) lot n° 2108.

Courtesy of Dr Lehner and Mrs Burel, University of Rouen Normandie, France

Application example

Total cell extract from Arabidopsis thaliana wild type (1) and cell extracts from different mutants defective in the complex N-glycan maturation pathway (2-5) (data not published yet).

Primary antibody has been used at 2 μ g/10 ml of incubation buffer. Detection has been done using ECL.



Dot blot reaction of anti-Fucose and anti-Xylose antibodies with various controls: Avidin (Fuc+/Xyl+), Fetuin (Fuc-/Xyl-), PLA2 (Fuc+/Xyl-) and Mur1-2 (Fuc-/Xyl+). 2 µl of each extract were spotted on a nitrocellulose membrane, placed on top of 2 WHATMAN filters (one soaked in TBS-T) and dried for 1.5 h at RT. The mem-brane was blocked for 30 min with 2% low-fat milk powder in TBS-T (0.1% TWEEN 20) and incubated with anti-Fucose(1) (<u>AS07 268</u>, 1:1000) or anti-Xylose(2) (<u>AS07 267</u>, 1:1000) for 30 min and then with secondary anti-rabbit(1:1000) antibody (ALP conjugated, recommended secondary antibody <u>AS09 607</u>). Membrane was washed with TBS-T 3 x 10 minutes before reaction development using alkaline phosphatase reagent BCIP®/NTB premixed solution (Sigma, Prod. No. B6404).

Please follow this link for a more detailed Dot-Blot protocol



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